ARTICLE



Heterozygous loss-of-function variants of *MEIS2* cause a triad of palatal defects, congenital heart defects, and intellectual disability

Rosalind Verheije¹ · Gabriel S. Kupchik² · Bertrand Isidor³,⁴ · Hester Y. Kroes⁵ · Sally Ann Lynch⁶ · Lara Hawkes⁵,⁵ · Maja Hempel⁰ · Bruce D. Gelb □¹0 · Jamal Ghoumid □¹¹ · Guylaine D'Amours¹² · Kate Chandler¹³ · Christèle Dubourg¹⁴ · Sara Loddo¹⁵ · Zeynep Tümer¹⁶ · Charles Shaw-Smith¹7 · Mathilde Nizon³ · Michael Shevell¹8 · Evelien Van Hoof¹ · Kwame Anyane-Yeboa¹⁰ · Gaetana Cerbone²⁰ · Jill Clayton-Smith¹³ · Benjamin Cogné³ · Pierre Corre²¹ · Anniek Corveleyn¹ · Marie De Borre¹ · Tina Duelund Hjortshøj¹⁶ · Mélanie Fradin²² · Marc Gewillig □²³ · Elizabeth Goldmuntz²⁴ · Greet Hens²⁵ · Emmanuelle Lemyre¹² · Hubert Journel²² · Usha Kini⁻,8 · Fanny Kortüm⁰ · Cedric Le Caignec³,⁴ · Antonio Novelli¹⁵ · Sylvie Odent²² · Florence Petit¹¹ · Anya Revah-Politi □²⁶ · Nicholas Stong □²⁶ · Tim M. Strom²,²,²8 · Ellen van Binsbergen⁵ · DDD study²⁰ · Koenraad Devriendt □¹ · Jeroen Breckpot¹

Received: 29 November 2017 / Revised: 3 July 2018 / Accepted: 15 July 2018 © European Society of Human Genetics 2018

Abstract

Deletions on chromosome 15q14 are a known chromosomal cause of cleft palate, typically co-occurring with intellectual disability, facial dysmorphism, and congenital heart defects. The identification of patients with loss-of-function variants in *MEIS2*, a gene within this deletion, suggests that these features are attributed to haploinsufficiency of *MEIS2*. To further delineate the phenotypic spectrum of the *MEIS2*-related syndrome, we collected 23 previously unreported patients with either a de novo sequence variant in *MEIS2* (9 patients), or a 15q14 microdeletion affecting *MEIS2* (14 patients). All but one de novo *MEIS2* variant were identified by whole-exome sequencing. One variant was found by targeted sequencing of *MEIS2* in a girl with a clinical suspicion of this syndrome. In addition to the triad of palatal defects, heart defects, and developmental delay, heterozygous loss of *MEIS2* results in recurrent facial features, including thin and arched eyebrows, short alae nasi, and thin vermillion. Genotype–phenotype comparison between patients with 15q14 deletions and patients with sequence variants or intragenic deletions within *MEIS2*, showed a higher prevalence of moderate-to-severe intellectual disability in the former group, advocating for an independent locus for psychomotor development neighboring *MEIS2*.

Introduction

Orofacial clefts are the most common craniofacial human birth defect, affecting the lip, both the lip, and the palate or the palate alone (CP). The spectrum of palatal clefts ranges from subclinical phenotypes like bifid uvula and high-arched palate, to submucous CP and velopharyngeal insufficiency, and to overt cleft palate. Most palatal clefts are isolated and sporadic, and are considered to have a

Electronic supplementary material The online version of this article (https://doi.org/10.1038/s41431-018-0281-5) contains supplementary material, which is available to authorized users.

Published online: 05 October 2018

Extended author information available on the last page of the article

multifactorial cause. Orofacial clefts, associated with developmental delay, dysmorphic features, or other major congenital anomalies, are defined syndromic. These mostly have a single genetic cause, either chromosomal or monogenic.

Deletions on chromosome 15q14 are a known chromosomal cause of cleft palate, typically co-occurring with intellectual disability (ID), facial dysmorphism and congenital heart defects (CHD) [1–9]. De novo loss-of-function variants in *MEIS2*, a gene within this region, were previously described in two patients with CP, ID, and CHD [10, 11]. Therefore, these features are attributed to haploinsufficiency of *MEIS2*. The *MEIS* genes belong to the three-amino-acid-loop extension (TALE) superfamily of homeobox (HOX) genes, highly preserved transcription factors consisting of a HOX with a helix-turn-helix structure of 60 amino acids, extended by a TALE between alpha



Fig. 1 Facial phenotypes from patients with de novo variants in *MEIS2* (patients 1–6, 8, and 9) and from patient K and L, who were identified with a de novo deletion on 15q14. Numbers or letters in the left upper corner refer to the patient numbering in Tables 1 and 2. Although facial phenotypes are variable and no clinically recognizable facial gestalt can be delineated, some recurrent features were noticeable, including thin, arched eyebrows, a metopic ridge, a thin vermillion, and short alae nasi. The bottom row shows evolution of facial features with age in patient K (from the age of 8 months to the age of 28 years). Note the thin, arched, and laterally displaced eyebrows, thin upper lip and broad nasal root and tip with short alae nasi, as well as facial coarsening with age

helices 1 and 2 in the homeodomain. MEIS2 most likely functions as a HOX co-factor, which binds to Pbx proteins and/or HOX proteins to form dimeric or trimeric complexes to enhance the specificity and affinity of DNA binding. MEIS2 is expressed during early fetal brain development in humans [12]. In zebrafish, Meis2 has been proven important in development of the mesencephalon [13], the craniofacial skeleton [14], and the heart [15, 16]. Conditional knockout of *Meis2* in developing murine neural crest cells results in abnormalities of the craniofacial skeleton, cranial nerves, and heart [17]. Results from animal models are in line with the triad of CHD, palatal defects and ID, previously observed in patients with variants or deletions of *MEIS2*.

To further delineate the phenotypic spectrum of this *MEIS2*-related syndrome, we collected 23 previously unreported patients either with a de novo variant in MEIS2 (patients 1–9), or with a 15q14 microdeletion encompassing this gene (patients A to N). Genotype–phenotype comparison was undertaken to identify recurrent features of the 15q14 deletion syndrome, which are not related to *MEIS2*, but rather to one of the neighboring genes.

Case reports

Detailed clinical reports of patients 1 to 9 are provided below. An overview of the features of the other patients (A to N) can be found in Table 2.

Patient 1

This boy is the second child of healthy, unrelated parents of North African descent. Familial history was negative with regard to developmental delay or congenital malformations. He was born at 39 weeks of gestation with weight 3.150 kg (25-50th centile), length 48 cm (10-25th centile) and head circumference 32 cm (below 3rd centile). At birth, cleft palate and a small perimembranous ventricular septal defect (VSD) were diagnosed. Hearing was normal. Clinical examination revealed proptosis and a downslant of the palpebral fissures. There was a mild eversion of the lower eyelids, fine arched eyebrows, and a small chin (Fig. 1). The metopic suture was prominent. At the age of 5 year 3 months, weight and height were at the 25th centile and head circumference was below the 3rd centile (-2.1 SD). The VSD had not closed spontaneously at that age. Psychomotor development was delayed. He walked at age 22 months. At age 3 years 5 months, nonverbal IQ was 64 (SON-R). Both fine and gross motor development were equivalent to the level of a child aged 2 years and 10 months. Speech was even more delayed, lagging behind more than 1.5 years.

Patient 2

This boy was born from healthy, unrelated parents of Caucasian origin. At birth he presented with cleft palate and

gastro-esophageal reflux. He developed conductive hearing loss and required bilateral hearing aids. He had sleep apnea, treated by nocturnal continuous positive airway pressure. This boy was referred to the genetics clinic because of developmental delay and behavioral problems. He started walking at the age of 18 months. His speech was moderately delayed with a predominant speech sound disorder with hypernasality. He was diagnosed with autism spectrum disorder and presented with temper tantrums, aggressive behavior, and short attention span. Brain MRI was normal, except for mild asymmetry of the lateral ventricles. At age 10 years 11 months, his height was 145.3 cm (50th centile), weight 40.5 kg (75th centile) and head circumference 56.5 cm (75-90th centile). Facial dysmorphic features included ptosis of the left eye, sagging of the lower eyelids, and square-shaped ear helices (Fig. 1).

Patient 3

Patient 3 is a 4-year-old boy, who was born full term as the first child of healthy, unrelated parents of Caucasian descent. The pregnancy was uncomplicated. He was described as an irritable baby and required syringed feeds for the first 3 days of life owing to poor suck. He has been managed for submucous cleft palate, hypermetropia, strabismus, and cryptorchidism. He was referred to the genetics clinic with autism spectrum disorder and developmental delay. He started walking independently from 30 months of age and had difficulties with co-ordination. He was not able to run or jump. He had 4–5 word sentences and understood simple instructions. His head circumference was 49.6 cm (5th centile). His weight and length were at the 25th centile. He presented with facial dysmorphism, featuring epicanthic folds, hypoplastic alae nasi, prominent ears, and a frontal cow lick (Fig. 1).

Patient 4

Patient 4 is a 20-year-old young woman of Caucasian descent, the second of four children. Pregnancy was uncomplicated. Delivery was at 42 weeks of gestation by cesarean section. Her birth weight was 4.5 kg. A median cleft palate was observed. After introduction of a special needs feeder, feeding was adequate. The cleft palate was surgically corrected at age 10 months. At the age of 6 months, she presented with limited movement with intermittent hyperextension of the trunk. Her muscular tone was normal. At age 12 months she was diagnosed with mild myopia with adequate vision. Hearing was intermittently compromised by middle ear infections. At age 14 months her motor and cognitive development was 6 months delayed. She could sit and walk unassisted, respectively, from the age of 2 and 3 years. She made sounds and spoke a few words at age 18 months; speech development was severely delayed. She attended a specialized school for children with compromised speech development. At age 20 years, however, speech is adequate, and in line with her cognitive development, which is mildly impaired.

Current biometric parameters are within normal range, with height being 173 cm (50th centile), weight 50 kg (10–25th centile), and head circumference 54.5 cm (25th centile). She has a broad forehead with bitemporal narrowing. Her limbs were normal, apart from diminished pronation and supination of the forearms. Brain MRI (at age 3 years), metabolic investigations, X-rays of the arms and hands (at age 2 years) were normal. Cardiologic investigations at age 20 years showed minimal billowing and insufficiency of the mitral valve.

Patient 5

This patient is a 14-year-old male of Mexican origin [18]. He was born at term following an uncomplicated pregnancy. Birth weight was 3.6 kg. He was noted to have cryptorchidism. Left testicle orchidopexy and inguinal hernia repair were performed at 1 year. Additional genitourinary surgeries included right-sided orchidopexy to correct retractile right testicle at 5 years and phimosis/ meatal stenosis repair at 6 years. Bifid uvula and possible submucosal cleft palate were identified at 16 months. Developmentally, he started walking with support at 16 months and independently at 18 months. He started speaking at 2 years and was able to speak in 2-3 word sentences by the age of three and a half. Brain and lumbar spine MRI and EEG completed at that time were unremarkable. At 2 years of age he was noted to have several dysmorphic features including prominent forehead, epicanthic folds, hypertelorism, long eyelashes with distichiasis, bulbous beaked nose with short alae nasi, bifid uvula, thin upper lip, retrognathia, short neck, hypoplastic right nipple, and fifth finger clinodactyly. Cardiac evaluation at that age was unremarkable and a murmur was deemed to be physiologic. He was noted to have precocious adrenarche and evaluated by an endocrinologist at the age of 5. HCG stimulation test showed borderline/low testosterone levels after stimulation. Myringotomy tubes were placed at 6 years owing to a history of recurrent ear infections and possible hearing loss. Ocular melanocytosis and iris nevus were noted on ophthalmology evaluation at 8 years. He was also noted to have "pre-glaucoma" at the age of 10 years old, for which he is being monitored. Most recent clinical evaluation at 13 years was significant for broad forehead, atypical medial eyebrow flare, distichiasis, micrognathia, mild posterior rotation of ears, and bifid uvula (Fig. 1). His head circumference was 57 cm (97th percentile), his height was 173 cm (98th percentile), and his weight was 72.3 kg (97th percentile).

Patient 6

Patient 6 is the first child of healthy, unrelated parents of Caucasian origin. She was born at term after an uncomplicated pregnancy. At birth she presented with a cleft palate. No associated anomalies were noted. She has fine arched eyebrows and hypoplastic alae nasi, but she was not considered dysmorphic (Fig. 1). She presented with mild developmental delay. She started walking at 25 months of age. Speech delay was noted which required speech therapy. Aged 5 years 8 months, her height was 101 cm (3rd centile), weight 13.5 kg (below 3rd centile (-2.5 SD)) and head circumference 49.5 cm (10th centile). Physical examination was normal. She exhibits learning difficulties.

Patient 7

This 18-year-old female is of Asian-Caucasian origin. She was born with a complex congenital heart defect: tetralogy of Fallot with an Ebstein's anomaly of the tricuspid valve. Early in life she had feeding difficulties, but neither palatal cleft, nor velopharyngeal insufficiency were reported. Psychomotor development was mildly delayed, requiring occupational therapies. At the age of 18 years, she was in the 11th grade of secondary school, without an individualized education program. She measured 160 cm (25th centile) and weighs 43.2 kg (3rd centile). She was not reported to be dysmorphic.

Patient 8

This girl was the first child of healthy, unrelated parents of Caucasian descent. The father had macrocephaly and the mother had a unilateral ear pit. Family history was otherwise unremarkable. The patient was born at 39 weeks of gestation. Birth weight was 2520 g (3rd centile), length was 48 cm (10-25th centile), and head circumference was 36 cm (90th centile). At birth, she presented with severe cyanosis, caused by an Ebstein's anomaly of the tricuspid valve with a hypoplastic right ventricle, a 7-mm perimembranous ventricular septum defect and a secundum atrial septum defect. In addition, she presented with a cleft of the palatum molle and a subtle notch in the hard palate. The congenital heart defect was treated surgically at the age of 8 months and, subsequently at the age of 3 years, to install a total cavopulmonary connection. The soft palate was surgically closed and ear tubes were placed at the age of 14 months. The first months of life were complicated by severe feeding problems, requiring tube feeding via a gastrostomy. This girl presented at the genetics clinic at 17 months, and was noted to have dysmorphic facial features including a high and broad forehead, fine arched eyebrows, hypoplastic alae nasi, short philtrum, and low-set dysplastic ears with a rightsided ear pit (Fig. 1). Hands and feet were normal, except for bilaterally overriding toes. She was delayed in attaining early developmental milestones. She walked independently from the age of 23 months. Until the age of 10 years, she needed speech therapy because of articulation difficulties and hypernasal speech. Currently, she attends a special needs school for children with learning problems. Her height and weight are within normal range, both mapping at the 25th centile. Her head circumference is at the 97th centile.

Patient 9

Patient 9 was the first child of a first and dizygotic twin pregnancy of healthy, unrelated parents of Caucasian descent. Family history was unremarkable. Prenatally an atrial and VSD were detected; a double bubble gave the suspicion of a duodenal stenosis. He was born prematurely by cesarean section at 32 weeks due to pathological Doppler velocimetry indicating a centralization of fetal circulation. Birth weight was 1340 g (10–25th centile), birth length 38 cm (5th centile), and occipitofrontal circumference at birth 29 cm (25th centile). Immediately after birth he required respiratory assistance. On the second day of life, a duodenal membrane causing stenosis was corrected. The atrial and ventricular septal defects were confirmed postnatally. In addition, stenosis of the left lower pulmonary vein was detected. The heart defect resulted in a heart failure, requiring closure of the septal defects at the age of 5 months. Bilateral inguinal hernia required treatment at the age of 7 months. The leading problem of this boy was a permanent respiratory insufficiency that could not be explained solely by his heart defect and was likely caused by malfunctioning pulmonary ventilation. The boy could not be weaned from respiratory assistance. Over the course of the disease he required artificial ventilation and a tracheostomy was implemented at the age of 11 months. In addition, he developed hypothyroidism, transitory pancreatitis, and nephrocalcinosis. He suffered from feeding difficulties, recurrent vomiting and failure to thrive. Repeatedly he had high temperatures without infections, probably owing to a disturbed central temperature regulation. His neurological development was poor. Brain ultrasound was normal at birth. Repeated brain imaging later in life showed bilateral ventriculomegaly and brain atrophy. At the age of 13 months his weight was 7.73 kg (< 3rd centile), length 65 cm (< 3rd centile (- 3.5 SD) and head circumference $44 \,\mathrm{cm}$ (< 3rd centile (-2.2 SD). He presented with profound developmental delay, severe muscular hypotonia of the trunk and hypertonia of the limbs. He had poor head control and he did not roll over. He did not interact and did not grasp. He showed a strained breathing and a restless behavior. Dysmorphic facial features included high and small forehead with a high frontal hairline, frontal bossing and temporal narrowing, short palpebral fissures, hypertelorism, a depressed nasal bridge with anteverted nares, full cheeks, and a small mouth (Fig. 1). Neck and limbs were short in comparison with the trunk. The boy died from respiratory insufficiency at the age 13 months.

Methods

Genomic DNA from the patients and their parents was extracted from peripheral leukocytes of ethylenediaminete-traacetic acid-treated blood according to standard methods. All procedures involving patients performed in this study were in accordance with the ethical standards of their respective institutional research committees. Written consent was obtained from all patients or from their legal representatives. Analysis is based on GRCh37/hg19 assembly. Exons are numbered according to NG_029108.1. Identified *MEIS2* variants were uploaded in the LOVD database (http://www.lovd.nl/MEIS2). Deletions, harboring *MEIS2* are available in the Decipher database (https://decipher.sanger.ac.uk/).

Copy Number analysis

For patients 1–8 chromosomal microarray analysis was performed, according to the manufacturer's instructions (supplementary methods). For patient 9, copy number variation was deduced from the whole-exome sequencing data (supplementary methods).

Next-generation sequencing (NGS)

Patients 1-7 and 9 were submitted to whole-exome sequencing (WES) for unexplained syndromic ID, CHD, and/or palatal defects. WES for patient 6 was restricted to the affected patient, followed by Sanger sequencing of potentially disease-causing variants in parental DNA. For all other patients NGS of parent-offspring trios was performed. For patient 2 and 3, WES was performed through the Deciphering Developmental Disorders (DDD) study, as described previously [19]. Patient 5 was enrolled in a research study at the Institute for Genomic Medicine at Columbia University (protocol AAAO8410). Patient 7 was recruited from the Pediatric Cardiac Genomics Consortium (PCGC) from the National Heart, Lung, and Blood Institute (NHLBI) and the Eunice Kennedy Shriver National Institute of Child Health and Human Development, and was analyzed as described by Homsy et al. [20]. A detailed overview of the applied NGS methods and variant annotation pipelines is provided as supplementary methods.

Sanger sequencing

For patient 8, targeted sequencing of all exons and exonintron boundaries of the *MEIS2* gene was performed based on phenotypical resemblance with previously described patients with *MEIS2* variants. Sanger sequencing was performed for the 12 coding exons and exon–intron boundaries of the *MEIS2* gene (NM_170674.4). Primers are available in supplementary table 1. The amplification products were purified, directly sequenced using universal primers with the BigDye terminator v3.1 Cycle Sequencing Kit (Applied Biosystems, Thermo Fischer Scientific, MA, USA), and analyzed on a 3730xl genetic analyzer (Applied Biosystems).

Sanger sequencing was applied in patients 1, 6, and 8, and their respective parents to confirm the de novo occurrence of the *MEIS2* variant in a heterozygous state.

Genotype-phenotype correlation with 15q14 deletion patients

PubMed abstracts were searched for reported patients with 15q14 deletions, which were identified or delineated by chromosome microarray analysis. The following deletions were excluded from further analysis: (1) known copy number polymorphisms (CNV reported in at least 1% of normal population databases), (2) deletions larger than 7 Mb in size, (3) deletions extending beyond chromosomal band 15q14, or (4) deletions not containing (at least one exon of) the gene *MEIS2*.

Subsequently, our locally curated CNV database and the publicly available database DECIPHER [21] were searched for unpublished patients with 15q14 deletions, in line with the inclusion criteria enlisted above. Phenotype requests were sent to the responsible clinician. Deletions for which no accurate phenotyping could be retrieved, were excluded from further analysis. Genotype–phenotype comparison was performed between patients with 15q14 deletions and patients with de novo variants in MEIS2, to find recurrent features in the deletion cohort, which could not be ascribed to haploinsufficiency of the gene *MEIS2*, but rather to one of the neighboring genes.

Results

Copy number analysis in patients 1-9

No known disease-causing copy number changes were identified in patients 1–8, undergoing chromosomal microarray analysis. In patient 5, SNP array analysis rendered a rare maternally inherited 6q25.2 duplication (chr6:g. (?_153413459)_(154286581_?)dup), which was considered

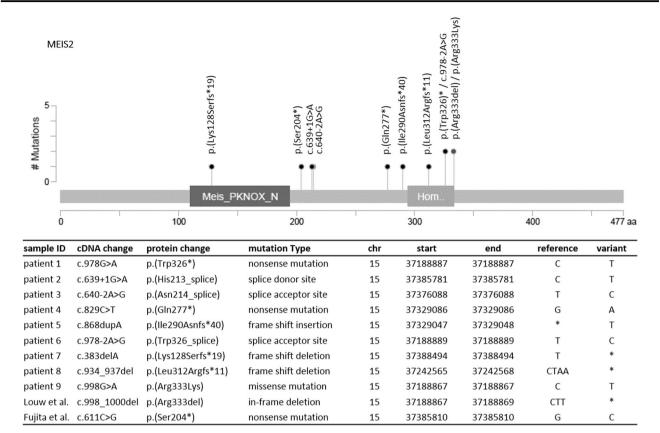


Fig. 2 Lollipop plot displaying novel and previously described de novo variants in *MEIS2*. Variants are plotted on a linear protein, using MutationMapper (cBioPortal-http://www.cbioportal.org/index.do) as described [34, 35]. Lollipops represent de novo protein changes in *MEIS2*. The dark and light gray box, respectively, represent the N-

terminal of homeobox Meis/PKNOX1 and the homeobox KN domain (or TALE-containing homeodomain). The p.(Arg333Lys) missense variant in patient 9 affects the same residue, that was found deleted in the patient reported by Louw et al. [10]

probably benign. Exome sequencing in patient 9 identified a de novo heterozygous deletion of 185 base pairs on chromosome 3p13 (chr3:g.(71179649_71179833)del), encompassing exon 1 of the less common *FOXP1* transcription variant NM_001244815.1.

Identification of de novo MEIS2 variants

A de novo variant in the gene *MEIS2* was identified in patients 1–9, either by WES, or by Sanger sequencing (three frameshift, two nonsense, three splice site, and one missense variant). Table 1 and Fig. 2 depict the positions of de novo variants in *MEIS2* in this cohort, complemented by two previously reported *MEIS2* variants [10, 11]. All de novo variants were absent in the 1000Genome (http://www.internationalgenome.org/), Exome Aggregation Consortium population databases (http://exac.broadinstitute.org/) and GnomAD databases (http://gnomad.broadinstitute.org/). The variants in patients 1–8 were predicted to cause protein truncation or nonsense-mediated decay, and therefore to cause loss-of-function. The missense variant in patient 9 was predicted to be damaging by in silico prediction

software. The recently developed prediction tool REVEL (rare exome variant ensemble learner) yielded a score of 0.838, corresponding to a high specificity for affecting function [22]. In addition, this patient was found to be compound heterozygous for two rare variants of unknown significance in the gene *LRP2*, c.8624G>A, p. (Arg2875His), and c.10424G>C, p.(Gly3475Ala) (c. [8624G>A];[10424G>C]; NM_004525.2/NP_004516.2). His unaffected twin brother was heterozygous for one of these variants. Patients 1–8 were not found to carry additional de novo, X-linked, compound heterozygous or homozygous variants, which could explain or contribute to their phenotype.

Clinical data from patients 1–9 and both previously published patients are summarized in Table 1. Key features of the *MEIS2*-related syndrome include palatal defects (9/10; 90%), ranging from bifid uvula to overt CP, ID (100%), CHD (7/11; 63%). Development is typically mildly delayed, with moderate-to-severe ID occurring in 4 out of 11 patients (36%). Three patients were reported with autism spectrum disorder. No recognizable facial gestalt could be ascribed to this syndrome. However, recurrent facial

Table 1 Clinical and molecular characteristics from patients harboring de novo variants in MEIS2 (NG_029108.1 (NM_170674.4))

	Fatient 3	Patient 4	Patient 5	Patient 6	Patient 7	Patient 8	Patient 9	Louw, 2015	Fujita, 2016	Total: 11 patients
000152867	#00152014	#00152015	#00152123	#0000152981	#0000152982	#0000152983	#0000152985	NA	NA	
lale	Male	Female	Male	Female	Female	Female	Male #	Female	Female	M/F: 5/6
.639+1G>A	c.640-2A>G	c.829C>T	c.868dupA	c.978-2A>G	c.383delA	c.934_937del	c.998G>A	c.998_1000del	c.611C>G	Inframe del
plice variant	Splice variant	p.(Gln277*)	p. (Ile290Asnfs*40)	Splice variant	p. (Lys128Serfs*19)	p. (Leu312Argfs*11)	p.(Arg333Lys)	p.(Arg333del)	p.(Ser204*)	1, stop codon 3, frameshift 3, Splice 3, missense 1
0 y 11 mo	4 y	20 y	13 y	5 y 8 mo	18 y 3 mo	11 y	1 y 1 mo	5 y 8 mo	2 y 9 mo	2 y - 20 y
Ţ.	Submucous CP	G.	Bifid uvula	CP	Not reported	Cleft soft palate	No	CP	CP	81%
Vormal	Normal	Mitral valve insufficiency	Normal	Normal	Tetralogy of Fallot, Ebstein malformation	Ebstein anomaly, VSD, ASD type 2	VSD, ASD, left pulmonary vein stenosis	ASD, VSD, LVOTO, CoA	ASD, VSD	63%
ysplastic ears	Prominent metopic ridge, portruding ears, epicanthic folds, hypoplastic alae nasi, prominent ears, cow's lick	Broad forehead with bitemporal narrowing	Broad forehead, flaring medial eyebronism, hypertelorism, districhiasis, thin upper lip, beaked nose, retrognathia	Fine arched eyebrows, hypoplastic alae nasi	Normal	High and broad forehead, fine arched eyebrows, hypoplastic alae nasi, short philtrum, low-set dysplastic ears, right-sided ear pit	Frontal bossing, high frontal hairline, temporal hairline, temporal narrowing, short palpebral fissures, hypertelorism, full cypes, low nasal bridge, anteverted nares, small mouth	Bitemporal narrowing, arched laterally extended eyebrows, deep-set eyes, thin upper lip	Large forehead, sparse eyebrows, mild mild urigonocephaly, deep-set eyes, low-set ears, full cheeks, thin upper lip	Bitemporal neartowing, prominent metopic ridge, fine arched laterally extended expedrows, hypoplastic alae nasi, thin upper lip (Fig. 1)
	49.6 cm (p5)	54.5 cm (p25)	57 cm (p97)	49.5 cm (p10)	NR	76d	44 cm (<p3)< td=""><td>50.7 cm (p50)</td><td>46.8 cm-1.5 SD</td><td>Variable,</td></p3)<>	50.7 cm (p50)	46.8 cm-1.5 SD	Variable,
45.3 cm 575)	16.4 kg (p25)	173 cm (p50)	173 cm (p98)	101 cm (p3)	160 cm (p25)	144.5 cm (p25)	65 cm (<p3)< td=""><td>108.5 cm (p10)</td><td>95 cm + 1 SD</td><td>failure to thrive in</td></p3)<>	108.5 cm (p10)	95 cm + 1 SD	failure to thrive in
0.5 kg (p75)	104.9 cm (p25)	50 kg (p10- 25)	72.3 kg (p97)	13.5 kg (<p3)< td=""><td>43.2 kg (p3)</td><td>33.2 kg (p25)</td><td>7.7 kg (p3)</td><td>15.8 kg (p3)</td><td>12.5 kg 0 SD</td><td>young children</td></p3)<>	43.2 kg (p3)	33.2 kg (p25)	7.7 kg (p3)	15.8 kg (p3)	12.5 kg 0 SD	young children
Aild-to- noderate ID	Mderate ID	Mild ID	Mild ID	Mild ID	Learning problems	Mild ID	Profound ID	Moderate ID	Severe ID	Mild (55%), moderate (27%), severe/ profound (18%)
8 months	30 months	3 y	18 months	25 months	NA	23 months	no walking	30 months	33 months	26 months (average)
	Autism	NR T	No	o O	NR	no	NA	Autism	NR T	Autism (60%)
	Feeding problems, strabismus, hypermetropia, cryptorchidism	Myopia (-4 D), reduced pronation underarms	Cryptorchidism, iris nevus, clino V, precocious adrenarche, scoliosis	Thin corpus callosum	Feeding problems, strabismus, congenital scoliosis	Feeding problems, oligodontia, overriding toes	Duodenal stenosis, feeding difficulties, hypothyroidism, inguinal hernia	Feeding problems, congenital lobar emphysema, II– III toe syndactyly	Feeding problems	Feeding problems in infancy
	#00152012 #0000152867 Male E.978G>A c.639+1G>A p.(Trp326*) Splice variant b.(Trp326*) Splice variant S y 3 mo 10 y 11 mo CP CP VSD Normal Prominent Prosis left eye, metopic dysplastic ears ridge. proptosis, eyes slant down, eyers slant down, eyers slant strated are nasi, arched age nasi, small chin 48 cm (cp3) 56.5 cm (p75) 107 cm (p25) 145.3 cm Mild ID Mild-to- moderate ID Mild-to- moderate ID No Autism, pervasive disorder NR NR	#00152014 Male c.640-2A>G Splice variant 4 y Submucous CP Normal Prominent in metopic ridge, protruding ears, epicamhie folds, hypoplastic alae nasi, prominent ears, cow's lick 104.9 cm (p.5) 16.4 kg (p.25) 104.9 cm (p.25) Mderate ID Mderate ID Autism Feeding problems, strabismus, hypermetropia, cryptorchidism	#00152014 #00152015 Male	#00152014 #00152015 Male C.640-2A>G C.829C>T Splice variant p.(Gln277*) 4 y Submucous CP CP Normal Miral valve insufficiency Prominent forehead with protruding ears, bitemporal epicanthic folds, narrowing hypoplastic alae nasi, prominent ears, cow's lick 49.6 cm (p5) 54.5 cm (p5) 16.4 kg (p25) 173 cm (p50) 104.9 cm (p25) 50 kg (p10-25) Mderate ID Mild ID Male ID Autism NR Feeding Myopia problemetropia, reduced hypermetropia, reduced hypermetropia, pronation cryptorchidism underarms	#00152014 #00152015 #00152123 Male C.640-2A>G C.829C>T C.868dupA Splice variant P.(Gln277*) Wilted valve Normal Normal Normal Mitral valve Prominent Broad Broad forehead, Jorehead with flaring medial protruding ears, bitemporal epicanthic folds, matrowing A9.6 cm (p5) 16.4 kg (p25) 173 cm (p25) 16.4 kg (p25) 173 cm (p26) 173 cm (p97) 16.4 kg (p25) Mid ID Mild ID Mild ID Mild ID Autism NR No No No No No No No No No	## ## ## ## ## ## ## ## ## ## ## ## ##	## ## ## ## ## ## ## ## ## ## ## ## ##	Water (AD) (150) (1	Maile Female Maile Female Female Female Maile Female Female Splice variant Prof. 13 10 10 10 10 10 10 10	Mail

Variant positions are according to genomic reference sequence Hg19. Patient 9 was found to be compound heterozygous for rare variants in the gene *LRP2*. In addition, a de novo deletion in the gene *FOXP1* was identified. *ASD* atrial septum defect, *CoA* aortic coarctation, *D* diopter, *ID* intellectual disability, *LOVD* Leiden open variation database: http://www.lovd.nl/MEIS2), *LVOTO* left ventricle outflow tract obstruction, *NA* not available, *NR* not reported, *p* centile, *SD* standard deviation, *VSD* ventricular septal defect

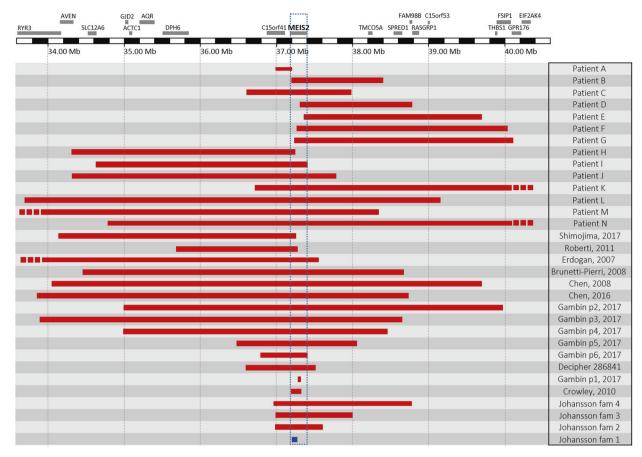


Fig. 3 Overview of 14 novel (patients A to N) and 17 previously reported deletions on chromosome 15q14, harboring at least one exon of the gene *MEIS2*. Deletions are depicted as red boxes. The intragenic

duplication, reported in family 1 by Johansson et al., is depicted as a blue box [6]. The region corresponding to the genomic position of the *MEIS2* gene is represented by a blue dotted box

features include thin, arched eyebrows, bitemporal narrowing, hypoplastic alae nasi, and a thin upper lip (Fig. 1).

Genotype-phenotype comparison with 15q14 deletion patients

Clinical and molecular details from 16 previously published families [1-9] and 15 novel patients (patients A to N, and Decipher patient 286841) with 15q14 deletions were collected (Fig. 3). Deletion sizes ranged from 123 kb to 6.97 Mb. Two deletions involved an intragenic deletion of MEIS2 [4, 9], whereas seven deletions only affected MEIS2 and its proximal neighbor C15orf41 (patient A and C, Decipher 286841, Johansson et al. [6], Gambin et al. [9]). Decipher patient 286841, as well as the six patients reported recently by Gambin et al. [9], were excluded from further genotype-phenotype analysis, as no sufficient clinical features could be retrieved (supplementary table 3). Twentyfive deletion carriers from 24 independent families were retained (patients A to N and 11 previously reported families [1–8]) (Table 2 and supplementary table 2). All 24 deletions affected the gene MEIS2, either completely (14/ 24), or partially (10/24). The deletion occurred de novo in 18 families and was inherited from an affected mother in one family (family 2 by Johansson et al. [6]). Inheritance was unknown in the five remaining families. Breakpoints of the deleted regions were unique, advocating for non-homologous end-joining as the underlying pathogenic mechanism of these 15q14 deletions. We also included one previously reported family with an intragenic tandem duplication in *MEIS2* with four affected family members (family 1 by Johansson et al. [6]). This duplication was predicted to cause loss-of function.

In three families copy number analysis rendered an additional rare variant elsewhere in the genome. Patient A had a 500 kb duplication on 5p15.31 (chr5:g.(?_8747001)_ (9247598_?)dup), and a 206 kb duplication on 5q15.2, harboring the gene *CTNND2* (chr5:g.(?_11298758)_ (11505375_?)dup). Both duplications were inherited from an unaffected mother. Intragenic deletions in *CTNND2* are considered a cause for ID [23]. It is currently unclear whether this partial *CTNND2* duplication occurred in tandem, and whether it contributed to the patient's phenotype. In patient C a de novo deletion on chromosome 12q23.3

 Table 2
 Molecular and clinical data of previously unpublished patients with 15q14 deletion

				,													
	Patient A	Patient B	Patient C	Patient D	Patient E	Patient F	Patient G	Patient H	Patient I	Patient J	Patient K	Patient L	Patient M	Patient N	This study: 14 novel families	Previously published families # (15 patients from 11 families)	Total: 25 patients from 24 deletion families, and 4 patients from 1 duplication family $(n = 29)$
Decipher ID		Patient 359852	Patient 339417	Patient 339417 Patient 251108 Patient 280740	8 Patient 280740	Patient 299872	nt 90	ent 259142	Patient 359853	Patient 257868	Patient 359854	ent 359855	ent 359856	ent 359858			
Sex	XX	XX	XX	XX						XY	X	XX	XX	XX	M/F = 9/5	M/F = 10/5	M/F = 19/5
15q14 del (hg19)	g.(? _36989551)_ (37184183_?) del		g.(? g.(?_36606036) g.(? _37196610)_ (37980027_?) _373 (38244056_?) del (387	. g.(? _37310981)_ (38781772_?) del		g.(? _37358935)37269953)_ (39695030_?) (40018834_?) del	g.(? _37231466)_ (40103247_?) del	g.(?_34308789) _(37231638_?) del	g.(?_34616375)_ (37401551_?)del	g.(?_34363958)_ (37792285_?)del	g.(?_36794841) _(41016404_?) del	g.(?_33707609)_ (39190567_?)del	g.(?_32260104) _(38338675_?) del	g.(? _34962033)_ (41938462_?) del	14 deletions	1 intragenic duplication, 10 deletions	1 intragenic duplication, 24 deletions (including 1 intragenic deletion)
Size (Mb)	194 kb	1.05 Mb	1.37 Mb	1.47 Mb	2.34 Mb	2.7 Mb	2.87 Mb	2.92 Mb	2.78 Mb	3.43 Mb	4.2 Mb	5.4 Mb	6.1 Mb	6.97 Mb	3.1 Mb (mean)	2.83 Mb (mean)	3 Mb (mean)
CGH platform	Agilent 60k	Signature Chip v2.0	Agilent 4x180k	Affymetrix SNP 6.0	Affymetrix CytoScan	Agilent 60k	OGT 8x60K v2.0	Agilent 60k	Nimblegen CGX-12 OGT 8 x 60k	OGT 8 x 60k	Agilent 4x180k	Agilent 4x180k	Agilent 60k	Nimblegen CGX-12	Variable	Variable	Variable
Inheriance	De по vо	De novo	De novo	Unknown	De novo	Unknown	Unknown	De поvо	De novo	De novo	De novo	De novo	De novo	Ŭnknown∗	10 de novo, 5 unknown	9 de novo, 1 familial deletion, 1 familial intragenic duplication	19 de novo, 1 familial deletion, 1 familial intragenic duplication
Other genetic defects	c chr5;g.(? _8747001) (9247598_?) dup; chr5;g. (?_11298758) _ (11505375_?) dup	N N	chrl2:g.(? 108249783) (108711392?) del	N.	X Z	Z Z	NR P	ž	Ä	X X	¥	N.	chrX :g.(? _122513198]_ (123056125_?) dup,mat	N.			
Age	13 y	3 y 6 mo	5 y	12 y	10 y 9 mo	13 y	NA	5 y 8 mo	5 y 6 mo	11 y	28 y	16 y	2 y 3 mo	5 y 6 mo	10 y (mean)	NA	NA
Palatal Defect	t Normal	Cleft palate	Normal	Bifid uvula	Normal	Bifid uvula, high-arched palate	Cleft palate	Normal	Cleft palate	Bifid uvula	Cleft palate	cleft palate	VPI	Cleft palate	10/14 (71%)	12/14 (85%)	22/28 (78%)
CHD	Normal	normal	Normal	Normal	Normal	Normal	Atrial septal defect	Normal	Normal	NR	Atrial AND ventricular septal defect	Mitral valve prolapse	PDA (surgery)	Tetralogy of Fallot	5/13 (38%)	7/15 (46%)	12/28 (43%)
Facial	Normal	Mild dysmorphic features	Prominent forehead, flat face, sparse eyebrows, epicanthic folds, hypertelorism, bulbous nasal tip	Protruding ears, high anterior hairline, long columella	Dysmorphic, wide mouth	Normal	Normal	Anteverted nares, asymmetric ears with abnormal belix	High forehead, hypertelorism, eyes slant down, broad philtrum, hypoplastic alae nasi, absent and andraisi groove, mild retrognathia	Convex nasal bridge, large prominent nose with beaked tip, short uptumed philtrum, thigh palate, prominent upper gums and central incisors		Large dysplastic ears, long and thin nose, hypoplastic alae nasi, low nasal niy, short philrum, dolichocephaly	Bitemporal narrowing, downslanting palpebral fissures, malar hypoplasia, short philtrum, small mouth	Short palpebral fissures, prominent masal bridge, pointed nasal tip, thin upper lip,	Mild and variable: bitemporal narrowing, downslanting papebra fissures, hypoplaxic alae nasi, dysplastic ears	Variable: bitemporal narrowing, pointed chin, beaked nose	Variable: bitemporal narrowing, downslamting palpebral fissures, hypoplastic alae nasi, dysplastic ears, pointed chin, beaked nose
Limb defects	Limb defects Hypoplasia of glenoid cavity	Normal	Clinodactyly V	Prominent fingertip pads	Pes planus, long and narrow feet	Normal	Normal	Normal	Normal	Arachnodactyly	Long fingers	Broad thumbs	Short hands and feet	o _N	No recurrent limb defects	No recurrent limb defects	No recurrent limb defects
Biometry	W: 52 kg (p75–90) L: 157 cm (p50–75) HC: 51 cm (<p3)< td=""><td>W: 11.6 kg (<p3) L: 90.6 cm (p3) HC: NA</p3) </td><td>W: 17kg (p25) L: 104cm (p10) HC: NA</td><td>W: 44.4kg (p75) L: 146.5 cm (p25) HC: 55.4 cm (p50)</td><td>W: 28 kg (p10) L: 136 cm (p25) HC: p3</td><td>W: 36 kg (p10) L: 152 cm (p25) HC: 54 cm (p25)</td><td>W: p9 L: p25 HC: p90</td><td>W: 13 kg (<p3) (<p3)="" (p3)<="" 49.5="" 99.5="" cm="" hc:="" l:="" td=""><td>W: 16 kg (p5) L: 107 cm (p25) HC: p50</td><td>W: 46.8 kg (p95) L: 154.3 cm (p98) HC: 55.7 cm (p75)</td><td>W: 67 kg (p75– 90) L: 159 cm (p25) HC: 52.3 cm (<p3)< td=""><td>W: 43.8 kg (p3) L: 162 cm (p5) HC: 55 cm (p10- 25)</td><td>W: p10 L: p50 HC: p10</td><td>W: 13 kg (<p3) (<p3)="" (<p3)<="" +hc:="" 47.8="" 99="" cm="" l:="" td=""><td>Short stature (3/ 13) microcephaly (5/12)</td><td>Short stature (3/6), microcephaly (3/5)</td><td>Short stature (6/19), microcephaly (8/17)</td></p3)></td></p3)<></td></p3)></td></p3)<>	W: 11.6 kg (<p3) L: 90.6 cm (p3) HC: NA</p3) 	W: 17kg (p25) L: 104cm (p10) HC: NA	W: 44.4kg (p75) L: 146.5 cm (p25) HC: 55.4 cm (p50)	W: 28 kg (p10) L: 136 cm (p25) HC: p3	W: 36 kg (p10) L: 152 cm (p25) HC: 54 cm (p25)	W: p9 L: p25 HC: p90	W: 13 kg (<p3) (<p3)="" (p3)<="" 49.5="" 99.5="" cm="" hc:="" l:="" td=""><td>W: 16 kg (p5) L: 107 cm (p25) HC: p50</td><td>W: 46.8 kg (p95) L: 154.3 cm (p98) HC: 55.7 cm (p75)</td><td>W: 67 kg (p75– 90) L: 159 cm (p25) HC: 52.3 cm (<p3)< td=""><td>W: 43.8 kg (p3) L: 162 cm (p5) HC: 55 cm (p10- 25)</td><td>W: p10 L: p50 HC: p10</td><td>W: 13 kg (<p3) (<p3)="" (<p3)<="" +hc:="" 47.8="" 99="" cm="" l:="" td=""><td>Short stature (3/ 13) microcephaly (5/12)</td><td>Short stature (3/6), microcephaly (3/5)</td><td>Short stature (6/19), microcephaly (8/17)</td></p3)></td></p3)<></td></p3)>	W: 16 kg (p5) L: 107 cm (p25) HC: p50	W: 46.8 kg (p95) L: 154.3 cm (p98) HC: 55.7 cm (p75)	W: 67 kg (p75– 90) L: 159 cm (p25) HC: 52.3 cm (<p3)< td=""><td>W: 43.8 kg (p3) L: 162 cm (p5) HC: 55 cm (p10- 25)</td><td>W: p10 L: p50 HC: p10</td><td>W: 13 kg (<p3) (<p3)="" (<p3)<="" +hc:="" 47.8="" 99="" cm="" l:="" td=""><td>Short stature (3/ 13) microcephaly (5/12)</td><td>Short stature (3/6), microcephaly (3/5)</td><td>Short stature (6/19), microcephaly (8/17)</td></p3)></td></p3)<>	W: 43.8 kg (p3) L: 162 cm (p5) HC: 55 cm (p10- 25)	W: p10 L: p50 HC: p10	W: 13 kg (<p3) (<p3)="" (<p3)<="" +hc:="" 47.8="" 99="" cm="" l:="" td=""><td>Short stature (3/ 13) microcephaly (5/12)</td><td>Short stature (3/6), microcephaly (3/5)</td><td>Short stature (6/19), microcephaly (8/17)</td></p3)>	Short stature (3/ 13) microcephaly (5/12)	Short stature (3/6), microcephaly (3/5)	Short stature (6/19), microcephaly (8/17)
Psychomotor development	. Moderate ID	Mild ID	Psychomotor delay	Intellectual disability	Intellectual disability	Moderate ID	Moderate ID	Intellectual disability	Motor delay, learning problems	Severe ID	Moderate ID	Moderate ID	moderate-to severe ID	Moderate-to- severe ID	Mild (2), moderate-severe (8), NFS (4)	Mild (9), moderate (2) NA (4)	Mild (11), moderate-severe (10), NA (4), NFS (4)
Walked at age	27 mo	18 mo	15 mo	24 mo	NA	14 mo	Not at 19 mo	30 mo	3 у	4 y	20 mo	34 mo	26 mo	Does not walk	14 mo-4 y (mean 14-36 mo 26 mo)		14-4 y
Behavior	Autism	NA	Autism	NA	NA	Normal	NA	Normal	ADD, anxiety	NA	Anxiety	N _o	autism	NA	Autism (3)	Autism (4)	Autism (7)

able 2	lable 2 (continued)	ed)															
	Patient A	Patient B	Patient A Patient B Patient C Patient D Patient E Patient F	Patient D	Patient E	Patient F	Patient G	Patient G Patient H	Patient I	Patient J	Patient K	Patient K Patient L	Patient M	Patient N	This study: 14 novel families	Previously published families # (15 patients from 11 families)	This study: 14 Previously Total: 25 patients novel families published from 24 deletion families, (15 families, and 4 parients from patients from 1 11 families) (15 families, and 4 patients from 1 1 1 families)
Other	Dyspraxia	°Z	Hypotonia, joint laxity, sacral dimple, paraumbilical hemia		Feeding Four large Large CAL problems, joint CAL spots, spots, laxity, CAL hypotonia, dyspraxia spot, broad suki cryptorchidism on MRI	Large CAL spots, dyspraxia	ои	Bitemporal depigmentation area, migraine- like headaches	Hypotonia, hyperlaxity	Severe myopia, joint laxity	Astignatism, hypermetropia, multiple naevi (no CAL spots)	nt Astigmatism. Joint laxity, Feeding Hyg hypemetropia, myopia, problems, skeep (loo nultiple naevi cryptorchidism, 4 disorder, GE mo (no CAL spots) CAL spots rellux, con hypotomic, hea hyperplaxity, fair hypophasic asti	Feeding problems, sleep disorder, GE reflux, hypotonia, hypotharity, hypoplastic nimples	Hypotonia (kower limb more affected), conductive hearing loss, fair skin, astigmatism	Hypotonia Hypotonia, pp (lower limb feeding more affected), difficulties, joint conductive hyperlaxity, CAL fearing loss, spots fair skin, astigmatism	_	CAL spots (SPREDI), feeding difficulties

ecently by Gambin et al. were excluded from further genotype—phenotype analysis, as no sufficient clinical features could be retrieved (supplementary table 3). CAL café-au-lait spots, HC head Father unavailable for testing, mother was normal. "An overview of previously published patients is found in supplementary table 2. Decipher patient 286841, as well as the six patients reported circumference, L length, NA not available, NR not reported, NFS intellectual disability not further specified, PDA patent ductus arteriosus, VPI velopharyngeal insufficiency, W weight was identified (chr12:g.(?_108249783)_(108711392_?)del). This deletion is absent from patient and normal population databases and affects two genes (WSCD2 and CMKLR1), which are not yet linked to developmental disorders. The duplication on Xq25, detected in patient M, harbors the genes THOC2 and XIAP, but not STAG2. Loss-of-function variants in THOC2 and XIAP are respectively causing X-linked recessive ID [24] and lymphoproliferative syndrome [25]. Duplications on Xq25, encompassing XIAP and STAG2, were considered disease-causing in two families with X-linked ID and facial dysmorphism, including malar hypoplasia and prognathism [26]. Although not comprising STAG2, this duplication might have contributed to the severe ID and malar hypoplasia in patient M.

Discussion

We contribute to the delineation of the MEIS2-related human phenotype by adding 23 novel patients either with de novo variants (patients 1-9), or with deletions of this gene (patients A to N). Novel de novo variants in MEIS2 included two nonsense variants (patient 1 and 4), three frameshift variants (patients 5, 6, and 7), three predicted splice variants (patients 2, 3, and 6) and one missense variant (patient 9). ExAc constraint scores of MEIS2 for loss-of-function and missense variants are respectively 0.99 and 3.20, indicating intolerance of MEIS2 towards loss-offunction and missense variants. The missense variant was found in a severely affected boy with profound ID and a complex heart defect. The question rises whether this severe and atypical presentation, including respiratory insufficiency, duodenal stenosis, and full cheeks, can solely be attributed to the p.(Arg333Lys) MEIS2 variant. Interestingly, this variant involves the same highly conserved Arg333 residue, affected by an in-frame deletion in the patient reported by Louw et al. [10]. As this residue is involved in the multimeric contact of MEIS2 [27], variants in this domain might exert a dominant negative effect by interference with the assembly of the multimeric compound, hereby contributing to the more severe phenotype observed in these two patients. Unfortunately, patient 9 died at the age of 13 months, and no tissue was available to perform functional analyses. Alternatively, his atypical features could be ascribed to multilocus genomic variation [28], as additionally he was (1) compound heterozygous for two rare variants in LRP2, and (2) he had a de novo intragenic 185-bp deletion of *FOXP1*. The patient's phenotype was not reminiscent of Donnai-Barrow (or faciooculoacousticorenal) syndrome [29] (OMIM:222448), which is caused by biallelic variants in LRP2. However, the FOXP1 deletion should be taken into account as loss-of-function variants and deletions of this gene are a known cause of global

developmental delay, severe speech delay and dysmorphic features, including frontal bossing, downslanting palpebral fissures, and a short, broad nose [30]. Interestingly, two features in our patient, duodenal atresia [31] and complex CHD [32], were previously attributed to deletions or variants of FOXP1. However, it is important to note that FOXP1 is subjected to alternative splicing and that the 185bp deletion is an intronic variant in the predominant expressed transcript and only affects an exon in a shorter alternative transcript. Therefore, the contributory effect of this *FOXP1* deletion on the phenotype remains speculative. Finally, premature birth at 31 weeks and complications from multiple invasive procedures might have contributed to worse outcome (e.g., respiratory insufficiency, inguinal hernia, small for gestational age, bitemporal narrowing, nephrocalcinosis, and pancreatitis) in patient 9. His dizygotic twin brother has mild developmental delay without any major congenital anomalies.

Recurrent features of 15q14 deletions, harboring at least one exon of MEIS2, include palatal and heart defects and ID, which are ascribed to haploinsufficiency of this gene. For phenotypic delineation of the 15q14 deletion syndrome, we retained 25 deletion carriers from 24 independent families. Palatal defects ranged from bifid uvula to overt cleft palate, and were present in 18 out of 24 deletion patients (75%), as well as in all four members of a family with an inherited intragenic duplication in MEIS2. Congenital heart defects were reported in 12 out of 24 deletion patients (50%), and mainly were septal defects. Given the high prevalence of heart defects in the de novo MEIS2 variant cohort, haploinsufficiency of MEIS2 was considered a sufficient cause of CHD in the deletion cohort. Whether neighboring genes on 15q14 contributed to or independently caused the genesis of CHD in the deletion cohort remains a subject of debate. In 14 patients, the 15q14 deletion comprised ACTC1, a known cause of septal heart defects and of autosomal dominant cardiomyopathy. The prevalence of CHD tends to be higher in patients with 15q14 deletions including ACTC1 (70% (7/10), compared with those without ACTC1 (39% (5/13)), but was not different from the CHD prevalence in the cohort of patients with de novo MEIS2 variants (63%). None of the 14 patients with haploinsufficiency of ACTC1 were diagnosed with dilated or hypertrophic cardiomyopathy. It is currently unclear whether whole-gene deletions of ACTC1 predispose to cardiomyopathy. Nevertheless, surveillance guidelines for patients with 15q14 deletions, encompassing ACTC1, should include cardiologic follow-up, even in the absence of CHD.

Developmental outcomes were available for 17 deletion patients and ranged from learning problems to severe ID. Moderate-to-severe ID was more prevalent in patients with 15q14 deletions, affecting *MEIS2* among other genes (10/17

(59%)), compared with patients with de novo variants [10, 11] or intragenic deletions/duplications [4, 6] in MEIS2 (4/ 14 (28%)), although no statistical significance could be reached (Fisher's exact test p = 0.14). Deletions extending distally from MEIS2 (beyond the gene SPRED1), as well as deletions reaching proximally into 15q13.3 can be associated with poor developmental outcome. Microcephaly is another recurrent feature of 15q14 deletions, that is seen less commonly in the cohort with de novo MEIS2 variants (2/11 (18%) versus 8/17 (47%), Fisher's exact test p =0.22). Although we presume that haploinsufficiency of neighboring genes on 15q14 independently affect brain growth and neurocognitive development, we were unable to identify additional candidate genes for ID or microcephaly on 15q14 based on the current data. Café-au-lait (CAL) spots were reported in patients D, E, F, and L, and in family 4 by Johansson et al. [6]. These deletions extend distally from MEIS2 and affect SPRED1, the causal gene for Legius syndrome, which is characterized by multiple café-au-lait macules, intertriginous freckling, lipomas, and learning disabilities [33] (OMIM: 611431). CAL spots were not reported in patients G, K, and N, and the patients described by Chen et al. [3, 7] and by Brunetti-Pieri et al. [2], although the deletions in these patients encompass SPRED1. This could be attributed to young age at diagnosis, variable expressivity, or inaccurate phenotyping.

Although no recognizable facial gestalt could be ascribed to this syndrome, one frameshift variant was found by targeted sequencing of *MEIS2* in a girl with a clinical suspicion of this syndrome. In addition to CHD and CP, this girl presented with thin, arched eyebrows, short alae nasi, and a thin vermillion, recurrent features in patients with deletions or loss-of-function variants of this gene (Fig. 1). In conclusion, deletions, indels or nucleotide variants in *MEIS2* should be considered in every patient presenting with syndromic palatal defects, particularly if associated with CHD, developmental delay, and facial dysmorphism.

Acknowledgements We thank the patients and their parents for their cooperation. J.B. is funded by a clinical research fund of the University Hospitals Leuven (Klinisch onderzoeksfond KOF Leuven). This study makes use of data generated by the DECIPHER community. A full list of centers who contributed to the generation of the data is available from https://decipher.sanger.ac.uk/ and via email from decipher@sanger.ac.uk. Funding for the project was provided by the Wellcome Trust. The DDD study presents independent research commissioned by the Health Innovation Challenge Fund [grant number HICF-1009-003], a parallel funding partnership between the Wellcome Trust and the Department of Health, and the Wellcome Trust Sanger Institute (grant number WT098051). The views expressed in this publication are those of the author(s) and not necessarily those of the Wellcome Trust or the Department of Health. The study has UK Research Ethics Committee approval (10/H0305/83, granted by the Cambridge South REC, and GEN/284/12 granted by the Republic of Ireland REC). The research team acknowledges the support of the National Institute for Health Research, through the Comprehensive Clinical Research Network. This study was supported by New York State Genetics Services Program C029418. This work grants U01HL131003, UM1HL098147, was supported by UM1HL098123, UM1HL128761, UM1HL128711, and UM1HL098162 in support of the Pediatric Cardiac Genomics Consortium from the National Heart, Lung, and Blood Institute and the Eunice Kennedy Shriver National Institute of Child Health and Human Development. The content of this article is solely the responsibility of the authors and does not necessarily represent the official views of the National Heart, Lung, and Blood Institute, Eunice Kennedy Shriver Institute of Child Health and Development, or the National Institutes of Health.

Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

References

- Erdogan F, Ullmann R, Chen W, et al. Characterization of a 5.3 Mb deletion in 15q14 by comparative genomic hybridization using a whole genome "tiling path" BAC array in a girl with heart defect, cleft palate, and developmental delay. Am J Med Genet A. 2007;143A:172-8.
- Brunetti-Pierri N, Sahoo T, Frioux S, et al. 15q13q14 deletions: phenotypic characterization and molecular delineation by comparative genomic hybridization. Am J Med Genet A. 2008;146A:1933–41.
- Chen CP, Lin SP, Tsai FJ, Chern SR, Lee CC, Wang W. A 5.6-Mb deletion in 15q14 in a boy with speech and language disorder, cleft palate, epilepsy, a ventricular septal defect, mental retardation and developmental delay. Eur J Med Genet. 2008;51: 368–72.
- Crowley MA, Conlin LK, Zackai EH, Deardorff MA, Thiel BD, Spinner NB. Further evidence for the possible role of MEIS2 in the development of cleft palate and cardiac septum. Am J Med Genet A. 2010;152A:1326–7.
- Roberti MC, Surace C, Digilio MC, et al. Complex chromosome rearrangements related 15q14 microdeletion plays a relevant role in phenotype expression and delineates a novel recurrent syndrome. Orphanet J Rare Dis. 2011;6:17.
- Johansson S, Berland S, Gradek GA, et al. Haploinsufficiency of MEIS2 is associated with orofacial clefting and learning disability. Am J Med Genet A. 2014;164A:1622-6.
- Chen CP, Chen CY, Chern SR, et al. Prenatal diagnosis and molecular cytogenetic characterization of a de novo 4.858-Mb microdeletion in 15q14 associated with ACTC1 and MEIS2 haploinsufficiency and tetralogy of Fallot. Taiwan J Obstet Gynecol. 2016;55:270-4.
- Shimojima K, Ondo Y, Okamoto N, Yamamoto T. A 15q14 microdeletion involving MEIS2 identified in a patient with autism spectrum disorder. Hum Genome Var. 2017;4:17029.
- Gambin T, Yuan B, Bi W, et al. Identification of novel candidate disease genes from de novo exonic copy number variants. Genome Med. 2017;9:83.
- Louw JJ, Corveleyn A, Jia Y, Hens G, Gewillig M, Devriendt K. MEIS2 involvement in cardiac development, cleft palate, and intellectual disability. Am J Med Genet A, 2015;167A:1142–6.
- Fujita A, Isidor B, Piloquet H, et al. De novo MEIS2 mutation causes syndromic developmental delay with persistent gastroesophageal reflux. J Hum Genet. 2016;61:835–8.
- Larsen KB, Lutterodt MC, Laursen H, et al. Spatiotemporal distribution of PAX6 and MEIS2 expression and total cell numbers

- in the ganglionic eminence in the early developing human forebrain. Dev Neurosci. 2010;32:149–62.
- Zerucha T, Prince VE. Cloning and developmental expression of a zebrafish meis2 homeobox gene. Mech Dev. 2001;102:247–50.
- Melvin VS, Feng W, Hernandez-Lagunas L, Artinger KB, Williams T. A morpholino-based screen to identify novel genes involved in craniofacial morphogenesis. Dev Dyn. 2013;242:817–31.
- Glickman NS, Yelon D. Cardiac development in zebrafish: coordination of form and function. Semin Cell Dev Biol. 2002;13:507–13.
- Paige SL, Thomas S, Stoick-Cooper CL, et al. A temporal chromatin signature in human embryonic stem cells identifies regulators of cardiac development. Cell. 2012;151:221–32.
- Machon O, Masek J, Machonova O, Krauss S, Kozmik Z. Meis2 is essential for cranial and cardiac neural crest development. BMC Dev Biol. 2015;15:40.
- Kupchik GS, Revah-Politi A, Stong N, Anyane-Yeboa K. Expanding genotype and phenotype of MEIS2-related neurocristopathy (P11.070B). Presented at the European Society of Human Genetics 28 May 2017, Copenhagen, Denmark.
- Wright CF, Fitzgerald TW, Jones WD, et al. Genetic diagnosis of developmental disorders in the DDD study: a scalable analysis of genome-wide research data. Lancet. 2015;385:1305–14.
- Homsy J, Zaidi S, Shen Y, et al. De novo mutations in congenital heart disease with neurodevelopmental and other congenital anomalies. Science. 2015;350:1262–6.
- Firth HV, Richards SM, Bevan AP, et al. DECIPHER: database of chromosomal imbalance and phenotype in humans using ensembl resources. Am J Hum Genet. 2009;84:524–33.
- 22. Ioannidis NM, Rothstein JH, Pejaver V, et al. REVEL: an ensemble method for predicting the pathogenicity of rare missense variants. Am J Hum Genet. 2016;99:877–85.
- Belcaro C, Dipresa S, Morini G, Pecile V, Skabar A, Fabretto A. CTNND2 deletion and intellectual disability. Gene. 2015;565:146–9.
- Kumar R, Corbett MA, van Bon BW, et al. THOC2 mutations implicate mRNA-export pathway in X-linked intellectual disability. Am J Hum Genet. 2015;97:302–10.
- Rigaud S, Fondaneche MC, Lambert N, et al. XIAP deficiency in humans causes an X-linked lymphoproliferative syndrome. Nature. 2006:444:110–4.
- Di Benedetto D, Musumeci SA, Avola E, et al. Definition of minimal duplicated region encompassing the XIAP and STAG2 genes in the Xq25 microduplication syndrome. Am J Med Genet A. 2014;164A:1923–30.
- 27. Krissinel E, Henrick K. Inference of macromolecular assemblies from crystalline state. J Mol Biol. 2007;372:774–97.
- Posey JE, Harel T, Liu P, et al. Resolution of Disease Phenotypes Resulting from Multilocus Genomic Variation. N Engl J Med. 2017;376:21–31.
- Donnai D, Barrow M. Diaphragmatic hernia, exomphalos, absent corpus callosum, hypertelorism, myopia, and sensorineural deafness: a newly recognized autosomal recessive disorder? Am J Med Genet. 1993;47:679–82.
- Le Fevre AK, Taylor S, Malek NH, et al. FOXP1 mutations cause intellectual disability and a recognizable phenotype. Am J Med Genet A. 2013;161A:3166–75.
- 31. Hamdan FF, Daoud H, Rochefort D, et al. De novo mutations in FOXP1 in cases with intellectual disability, autism, and language impairment. Am J Hum Genet. 2010;87:671–8.
- Chang SW, Mislankar M, Misra C, et al. Genetic abnormalities in FOXP1 are associated with congenital heart defects. Hum Mutat. 2013;34:1226–30.
- Brems H, Chmara M, Sahbatou M, et al. Germline loss-offunction mutations in SPRED1 cause a neurofibromatosis 1-like phenotype. Nat Genet. 2007;39:1120–6.

- Gao J, Aksoy BA, Dogrusoz U, et al. Integrative analysis of complex cancer genomics and clinical profiles using the cBio-Portal. Sci Signal. 2013;6:p11.
- 35. Cerami E, Gao J, Dogrusoz U, et al. The cBio cancer genomics portal: an open platform for exploring multidimensional cancer genomics data. Cancer Discov. 2012;2:401–4.

Affiliations

Rosalind Verheije¹ · Gabriel S. Kupchik² · Bertrand Isidor³,⁴ · Hester Y. Kroes⁵ · Sally Ann Lynch⁶ · Lara Hawkes⁵,⁵ · Maja Hempel⁰ · Bruce D. Gelb □¹0 · Jamal Ghoumid □¹¹ · Guylaine D'Amours¹² · Kate Chandler¹³ · Christèle Dubourg¹⁴ · Sara Loddo¹⁵ · Zeynep Tümer¹⁶ · Charles Shaw-Smith¹7 · Mathilde Nizon³ · Michael Shevell¹8 · Evelien Van Hoof¹ · Kwame Anyane-Yeboa¹⁰ · Gaetana Cerbone²⁰ · Jill Clayton-Smith¹³ · Benjamin Cogné³ · Pierre Corre²¹ · Anniek Corveleyn¹ · Marie De Borre¹ · Tina Duelund Hjortshøj¹⁶ · Mélanie Fradin²² · Marc Gewillig □²³ · Elizabeth Goldmuntz²⁴ · Greet Hens²⁵ · Emmanuelle Lemyre¹² · Hubert Journel²² · Usha Kini⁻,⁵ · Fanny Kortüm⁰ · Cedric Le Caignec³,⁴ · Antonio Novelli¹⁵ · Sylvie Odent²² · Florence Petit¹¹ · Anya Revah-Politi □²⁶ · Nicholas Stong □²⁶ · Tim M. Strom²,²,² · Ellen van Binsbergen⁵ · DDD study²⁰ · Koenraad Devriendt □¹ · Jeroen Breckpot¹

- Center for Human Genetics, Catholic University Leuven, Leuven, Belgium
- Division of Medical Genetics, Infants and Children's Hospital of Brooklyn, Maimonides Medical Center, Brooklyn, NY, USA
- ³ CHU Nantes, Service de génétique médicale, Nantes, France
- ⁴ INSERM UMR 1238, Sarcomes osseux et remodelage des tissus calcifiés, Université Bretagne Loire, Nantes, France
- Department of Medical Genetics, University Medical Center Utrecht, Utrecht, Netherlands
- Departments of Clinical Genetics, Children's University Hospital Temple Street, Dublin, Ireland
- Oxford Centre for Genomic Medicine, Oxford University Hospitals NHS Foundation Trust, Oxford, UK
- Spires Cleft Service, Oxford University Hospitals NHS Foundation Trust, Oxford, UK
- ⁹ Institute of Human Genetics, University Medical Center Hamburg-Eppendorf, Hamburg, Germany
- The Mindich Child Health and Development Institute and the Departments of Pediatrics and Genetics and Genomic Sciences, Icahn School of Medicine at Mount Sinai, New York, New York, USA
- Service de Génétique Clinique, Hôpital Jeanne de Flandre, CHU Lille, Lille, France
- Service de génétique médicale, CHU Sainte-Justine, Département de Pédiatrie, Université de Montréal, Montréal, QC, Canada
- Manchester Centre for Genomic Medicine, Manchester University Hospitals NHS Foundation Trust, Manchester Academic Health Science Centre, Manchester, UK
- Laboratoire de Génétique Moléculaire, CHU Pontchaillou, Rennes, France
- Laboratory of Medical Genetics, Bambino Gesu Children's

- Hospital, IRCCS, Rome, Italy
- Applied Human Molecular Genetics, Kennedy Center, Department of Clinical Genetics, Copenhagen University Hospital Rigshospitalet, Glostrup, Denmark
- ¹⁷ Institute of Biomedical & Clinical Science, University of Exeter Medical School, Exeter, UK
- Department of Pediatrics, Faculty of Medicine, McGill University, Montreal, Ouebec, Canada
- Division of Clinical Genetics, Department of Pediatrics, Columbia University Medical Center, New York, NY, USA
- Division of Medical Genetics, "S.G. Moscati" Hospital, Avellino, Italy
- ²¹ Service de Stomatologie, CHU Nantes, Nantes, France
- Service de Génétique Médicale, Centre de Reference Anomalies du Développement, CHU Rennes, Rennes, France
- Pediatric and Congenital Cardiology, UZ Leuven, Leuven, Belgium
- Division of Cardiology, Children's Hospital of Philadelphia, Philadelphia, PA, USA
- Department of Otorhinolaryngology-Head and Neck Surgery, University Hospitals Leuven, Leuven, Belgium
- ²⁶ Institute for Genomic Medicine, Columbia University Medical Center, New York, NY, USA
- Institute for Human Genetics, Helmholtz Zentrum München, Neuherberg, Germany
- Institute for Human Genetics, Technische Universität Munchen, Munich, Germany
- DDD Study, Wellcome Trust Sanger Institute, Hinxton, Cambridge, UK