



Red blood cell algorithms in the diagnosis of red blood cell diseases and anaemia

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What is anaemia?



- Major public health problem: **25%** of the entire world population
- WHO: "decrease in the number of red blood cells or Hb concentration"
- Persistent decrease in O2 supply \rightarrow irreversible organ dysfunction

Detecting anaemia and the underlying cause in an early phase is highly demanded!

Different types of anaemia



Microcytic anaemia MCV < 80 fL	Normocytic anaemia MCV: 80-100 fL	Macrocytic anaemia MCV > 100 fL
Iron deficiency	Acute bleeding	VitB12/folate decifiency
Thalassaemia	Haemolytic anaemia:	Drug-induced
Chronic bleeding	 Enzyme defects (PKD, G6PD) 	MDS
Haemolytic anaemia (HS)	Hemoglobinopathies	Liver disease or alcohol use
Less common:	 Membrane defects (HS, PNH) Auto- and alloantibodies (AIHA) 	Hypothyroidism
Chronic/infectious disease, MDS	 Microangiopathic (TTP, aHUS) Drug-induced Infections Toxic substances Metabolic disorders Chronic/infectious disease Hypersplenism 	
	Less common: Iron deficiency, VitB12/folate, MDS	

Screening algorithms

- Facilitate the differential diagnosis of anaemia Favor a timely diagnosis and treatment
- Based on **RBC parameters**
 - Conventional parameters (e.g., MCH, MCV, ...)
 - Research parameters (e.g., microRBC, Hypo-He,...)
- Confirmatory tests are required
- Adequate screening algorithm: high sensitivity and specificity
- Sysmex RBC suspect flags: "HGB defect?", "Fragments?", "Iron deficiency?"





Research questions





Which algorithms based on red blood cell (research) parameters are reported in literature to facilitate the diagnostic work-up of red blood cell diseases and anaemia?



Which algorithms could be useful as a **screening tool** for distinct types of red blood cell diseases and **anemia in our laboratory**? What is the **diagnostic performance** of these algorithms in our laboratory?

With a focus on ...



Macrocytic anaemia

MCV > 100 fL

VitB12/folate decifiency

- 1. β-thalassaemia
- 2. Iron deficiency anaemia
- 3. Hereditary spherocytosis
- 4. Pyruvate kinase deficiency

Haemolytic anaemia Drug-induced Thalassaemia (PKD, G6PD) Enzyme defect MDS Chrome bleeding Haemolytic anaemia (HS) Liver disease or alcohol use Hemoglobinop Membrane defect Hypothyroidism Less common: Microangiopathic (TTP, aHUS Chronic/infectious disease, MDS Infections Toxic substances Metabolic disorders Chronic/infectious disease Hypersplenism Less common:

Iron deficiency, VitB12/folate, MDS

Acute bleeding

Normocytic anaemia

MCV: 80-100 fL

Microcytic anaemia

MCV < 80 fL

Iron deficiency

5. Microangiopathic haemolytic anaemia

RBC research parameters

→ Sysmex XN-9100









- Minor, major or intermedia
 - > based on the genotype, the extent of anaemia, and the clinical symptoms
- Production of β -globin chains impaired + excess of α -globin chains:
 - increase in HbA2 (α2δ2) levels (> 3.2%)
- Accumulation of **unstable α-globin chain tetramers** in erythroid cells
 - > ineffective erythropoiesis and peripheral haemolysis







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 - > ineffective erythropoiesis and peripheral haemolysis
- Treatment:
 - management of anaemia
 - haematopoietic stem-cell transplantation (if severe disease)



β-thalassaemia: algorithms



- Microcytic, hypochromic anaemia (~Iron deficiency anaemia)
- Since 1973...

Reference	Formula	Cut-off for thalassaemia
Ehsani et al, 2009	MCV – (10*RBC)	<15
England et al,1973	MCV - RBC - 5*Hb - 3.4	<0
Green and King, 1989	MCV ² * RDW/100 * Hb	<65
Mentzer, 1973	MCV/RBC	<13
Ricerca et al, 1987	RDW/RBC	<4.4
Shine and Lal, 1977	MCV ² * MCH * 0.01	<1530
Sirdah et al, 2008	MCV – RBC – 3 * Hb	<27
Srivastava and Bevington, 1973	MCH/RBC	<3.8
Urrechaga et al, 2008	M/H (=microRBC/Hypo-He)	>3.7
Urrechaga et al, 2011*	MicroRBC – Hypo-He – RDW-CV	>-5.1 or >-7.6

*β-thalassaemia patients: n=270, IDA: n=250, apparently healthy subjects: n=90











Adam et al, LHUB-ULB (Belgium) IDA (n=56), BTM (n=68), HS (n=14), heterozygote hemoglobinopathies (n=24) and others (n=118) IDA 🗖 56 BTM 📒 68 HGB HTZ 24 Noeud : 1 Good classification rate: 86.6%. Taille : 280 HS 14 %:100 OTHERS 118 Pureté(%): 42.1 RET-HE **β-thalassaemia** Sensitivity: 92.6% IDA 🗧 49 IDA 7 ≤ 25.6 > 25.6 BTM 67 BTM 1 HGB HTZ 2 HGB HTZ 22 Specificity: 94.8% Noeud : 2 Noeud:3 Taille : 121 Taille : 159 HS 0 HS 14 %:43.2 %:56.8 OTHERS 3 OTHERS 115 Pureté(%) : 55.4 Pureté(%):72.3 %MicroR/%Hypo-He %MicroR IDA 📘 IDA 8 IDA 0 IDA 7 > 6.6 ≤ 2.25 > 2.25 ≤ 6.6 BTM 4 BTM 63 BTM BTM 1 Noeud : 7 Noeud : 4 HGB HTZ 1 Noeud:5 HGB HTZ 1 Noeud : 6 HGB HTZ 3 HGB HTZ 19 Taille : 47 Taille : 126 Taille : 33 HS 0 Taille : 74 HS 0 HS 11 HS 3 %:16.8 %:26.4 %:45 %:11.8 OTHERS OTHERS 2 OTHERS 112 OTHERS 3 Pureté(%): 87.2 Pureté(%): 85.1 Pureté(%):88.9 Pureté(%) : 57.6 RET/IRF BTM IDA HGB HTZ IDA IDA 0 ≤ 9.97 > 9.97 BTM 0 BTM HGB HTZ 3 HGB HTZ Noeud : 12 Noeud:13 0 Taille : 103 Taille : 23 HS 0 HS 11 %:36.8 %:8.2 IDA OTHERS 12 Pureté(%):97.1 OTHERS Pureté(%) : 52.2 BTM HGB HTZ HS OTHERS HS OTHERS







Diagnostic performance

ROC analysis study population [MicroRBC - Hypo-He - RDW-CV] (β-thalassaemia *n*=18, controls *n*=289)

> Total number of samples **indicated as positive** using the distinct algorithms

02/02/2021 – 02/05/2021	Total number of samples	M/H [MicroRBC/Hypo- He] > 3.7	Urrechaga [MicroRBC- Hypo-He-RDW- CV] > -5.1	Urrechaga [MicroRBC- Hypo-He-RDW- CV] > -7.6	Adam Ret-He ≤ 25.6 AND [MicroRBC/Hypo- He] > 2.25	Sysmex flag "Hgb defect?"
Total population (< 18 and ≥ 18 years)	15376	7153 (47%)	195 (1%)	409 (3%)	181 (1%)	39 (0.3%)
Adult population (≥ 18 years)	11567	5050 (44%)	34 (0.3%)	71 (0.6%)	89 (0.8%)	4 (0.03%)

 \rightarrow ± 2 samples/day

β-thalassaemia

Implementation – to do:

- [MicroRBC Hypo-He RDW-CV] > -5.1 (Urrechaga et al, 2011)
- Samples will be forwarded to a **separate worklist** within the LIS.
- The clinical biologist checks the list **on a daily basis**.
- If suspicious for β-thalassaemia (based on the patient's history, blood parameters and clinical file), a **reflex test** for hemoglobin electrophoresis should be performed or, if sample volume is too low, a **comment** should be added to the lab report.

- Barre

- Most common cause of anaemia
- Driven by insufficient iron intake
 - by malnutrition
 - > during periods in life with higher demand i.e. pregnancy or growth
- Bone marrow iron stores using Perls' Prussian blue staining
 = gold standard, but requires invasive procedure
- Biochemical parameters: serum ferritin levels, transferrin levels/saturation (TSAT) and soluble transferrin receptors
 - \rightarrow acute phase reactants!

Reticulocytes survive 1 to 3 days in the peripheral blood circulation
 → Ret-He accurately reflects the actual iron availability in the bone marrow

- Ret-He is also decreased in patients
 - with β-thalassaemia
 - with ACD/acute inflammation
 (= functional iron deficit mainly due to hepcidin-25)

Discriminating formulas or algorithms!

Iron deficiency anaemia

Nivaggioni et al, 2020 IDA (n = 120), heterozygous haemoglobinopathy (n = 92), SCD (n = 56), HS (n = 18), other patients (n = 7931)

Adam et al, LHUB-ULB (Belgium)

IDA (n=56), BTM (n=68), HS (n=14), heterozygous hemoglobinopathy (n=24) and others (n=118)

IDA: retrospective study

IDA: retrospective study

Diagnostic performance

IDA: retrospective study

No implementation:

- sensitivity and specificity adequate
- PPV is rather low (29.4%)
- ± 14 positive samples per day \rightarrow only 4 with IDA

- Haemolytic anaemia with a normocytic appearance
- Highest prevalence of HS is in **Northern Europe** (1 in 2000)
- Mutations in genes encoding distinct proteins of the RBC membrane or cytoskeleton
 - > elastic deformability is affected
 - shortened lifespan of spherocytic RBC

- Laboratory tests:
 - > the **osmotic fragility** test: low sensitivity and specificity
 - > the observation of spherocytes on a **blood smear**.
 - > The flow cytometry-based eosin-5'-maleimide (EMA)-binding test
 - > the cryohaemolysis test
 - > SDS-PAGE
 - > ektacytometry
 - > molecular testing
- Specific limitations:
 - > low sensitivity and specificity, time-consuming procedures, or scarce availability
 - screening tools could have an added value

> Screening algorithms

Parameters	Bobée et al, 2018 (n= 47 HS, 17 PKD, 118 other (e.g. sickle cell, thalassaemia) and 489 routine samples)	Adam et al, 2020 (n=29 HS, 58 non-HS)	
Reticulocytes	> 80*10^9/L	> 101*10^9/L	
Ret/IRF	> 9.1	> 8.35	
Hb < 12 g/dL: MicroRBC	> 2.2%	/	
Hb < 12 g/dL: MicroRBC/Hypo-He	/	≥ 1.6	
Hb > 12 g/dL: MicroRBC/Hypo-He	≥ 3.5	≥ 3.8	
Sensitivity	100%	100%	
Specificity	92.1%	97.3%	

- Interpretation of the reticulocyte scattergram in the presence of the 'abnormal RET scattergram' analyser flag
 - rejection if the reticulocytes are not properly gated and/or separated from the RBC gate
 - > "unmeasurable due to analytical interference"
- Spherocytes contribute to an abnormal RET scattergram
 - Algorithms using parameters extracted from the LIS will only select samples without the morphological presence of spherocytes

Smear review of 28 samples with "analytical interference"

- > three patients with known auto-immune hemolytic anaemia
- three patients with known HS

Implementation – to do:

- lab technicians should be trained to recognize **RET scattergram patterns** indicative of spherocytes
- a **smear review** should be performed on suspicious samples and in case spherocytes are present, a **comment should be added to the lab report**

- Haemolytic anaemia with a normocytic appearance
- 1/20 000 in the general Caucasian population

PK generates 50% of the RBC total ATP
 → decreased ATP levels reduce the RBC lifespan

• autosomal recessive disorder often lacking a prominent parental history

- Laboratory tests:
 - > measurement of **PK enzyme activity** using spectrophotometric analysis
 - > *PK-LR* **molecular** testing
 - combining both tests is recommended for a definite diagnosis of PKD and adequate patient management

→ a screening algorithm based on RBC parameters could facilitate the diagnosis of PKD and increase awareness to the clinicians

> Screening algorithm for PKD

Parameters	Bobée et al, 2018 (n= 47 HS, 17 PKD, 118 other (e.g. sickle cell, thalassaemia) and 489 routine samples)		
Reticulocytes	> 150*10^9/L		
Ret/IRF	> 9.5		
MicroR	< 5.5%		
MicroRBC/Hypo-He	< 6.0		
Sensitivity	100%		
Specificity	96.5%		

Prospective study:

10 samples indicated as positive by the algorithm, with anaemia and without known haematological disorders (eg., acute leukemia)

Patient n°	Age (y)	Gender (m/f)	Hb (g/dL)	PK activity (U/g Hb)
1	33	m	8,7	4,8
2	64	f	13,0	6,6
3	90	m	11,0	10,9
4	68	m	7,3	10,0
5	65	f	9,2	7,2
6	38	f	8,9	8,3
7	29	f	10,9	7,5
8	76	m	11,8	6,7
9	91	f	11,5	6,1
10	41	f	10,9	0,2

PKD: retrospective study

Retrospective study:

01/10/2021 - 02/05/2021

- 36 routine samples were analyzed for both PK activity and reticulocyte parameters (same prescription): normal PK activity (> 3.8 U/g Hb)
- > non-suspicious for PKD based on the algorithm

02/02/2021 - 02/05/2021

92 out of the 16550 (0.6%) samples are suspicious for PKD according to the algorithm (± 1 sample/day)

PKD: retrospective study

Implementation – to do:

- Positive samples will be forwarded to a separate worklist in the LIS
- The clinical biologist will check the list on a daily basis
- If appropriate, a **comment** will be added to the lab report
 - the clinician may be guided swiftly in the right direction and should assess whether the measurement of PK activity is appropriate or not in a particular patient

Microangiopathic haemolytic anaemia

- Non-immune intravascular haemolysis
 - structural damage to the RBC membrane
 - formation of red cell fragments or schistocytes
- Shear stress

- > defective mechanical hearth valves, abnormalities in the microvasculature
- Thrombotic microangiopathies (TMA)
 - > TTP, HUS, drug-induced TMA, and complement-mediated TMA
- Other conditions
 - pregnancy-associated syndromes, severe hypertension, systemic infections and malignancies, ...
- TMAs are **life-threatening** medical urgencies: fast diagnosis is important!

Microangiopathic haemolytic anaemia

- Golden standard: smear review
 - ICSH: a schistocyte count > 1% on peripheral blood smear has a definite clinical value for the diagnosis of TMA in the absence of additional severe red cell shape abnormalities
 - inter-observer variations and time-consuming
- Haematology analyzers: automated count FRC
 - sensitivity and specificity remains ambiguous
- Sysmex suspect RBC flag: "Fragments?"
 - combination of FRC, RDW-SD, PLT upper discriminator, MCV, RBC lower discriminator, MCHC, MP

➤ 11/03/2021 and 10/05/2021: n=11180 samples

> 11/03/2021 and 10/05/2021: *n*=571 samples

Microscopic assessment: the maximum number of schistocytes counted	Score
≤ 6 schistocytes/ 1000 RBC	0
7-10 schistocytes/ 1000 RBC	1
11-20 schistocytes/ 1000 RBC	2
> 20 schistocytes/ 1000 RBC	3

> ROC analysis

Schistocytosis: schistocyte score ≥ 2

Pre-condition	Number of samples with score ≥ 2	Number of samples with score < 2	AUC
1. All samples	25	546	0.8286
2. MCH > 27 pg	23	487	0.8519
3. MCH > 27 pg and Hypo-He < 1.2%	9	382	0.9254

➤ Diagnostic performance (Schistocytosis: schistocyte score ≥ 2)

- ▶ RBC suspect flag "Fragments?":
 Only 10 out of 25 (40%) with score ≥ 2
 → insufficient
- > Number of **selected samples** using the distinct FRC% cut-offs

Pre-condition	Cut-off	Number of samples
1. All samples (n=11180)	> 0.78	2328 (21%, ± 39 samples/day)
2. MCH > 27 pg (n=10002)	> 0.78	1731 (17%, ± 29 samples/day)
3. MCH > 27 pg and Hypo-He < 1.2% (n=8410)	> 0.78	352 (4%, ± 6 samples/day)
1. All samples (n=11180)	> 0.91	2100 (19%, ± 35 samples/day)
2. MCH > 27 pg (n=10002)	> 0.91	1534 (15%, ± 26 samples/day)
3. MCH > 27 pg and Hypo-He < 1.2% (n=8410)	> 0.91	220 (3%, ± 4 samples/day)
1. All samples (n=11180)	> 1.00	1949 (17%, ± 33 samples/day)
2. MCH > 27 pg (n=10002)	> 1.00	1400 (14%, ± 23 samples/day)
3. MCH > 27 pg and Hypo-He < 1.2% (n=8410)	> 1.00	154 (2%, ± 3 samples/day)

No implementation as a screening tool:

- Specificity too low
- Too much samples that would have to be screened (35 samples/day)
- FRC% may be used supplementary to the microscopic assessment of schistocytosis. This is in line with the current ICSH guidelines on the assessment of schistocytosis.

- **Implementation** of the following screening algorithms:
 - Formula of Urrechaga et al to screen for β-thalassaemia: [MicroRBC – Hypo-He – RDW-CV] > -5.1
 - Formula of Bobée et al to screen for PKD: [RET > 150*109/L, Ret/IRF > 9.5, MicroRBC < 5.5% and MicroRBC/Hypo-He] < 6.0</p>
- Lab technicians should be trained to recognize the RET scattergram pattern indicative of **HS**.
- **Evaluation** of the usefulness of these screening tools a few months and/or one year following the implementation.

- This study showed the potential usefulness of the implementation of three screening tools in the context of anaemia and RBC pathologies in the clinical laboratory of UZ Leuven
- Our results highlight the importance of evaluating the diagnostic performance of published screening algorithms using lab-specific sample populations and haematology analyzers.

References

1. De Benoist B, McLean E, Egli I, Cogswell M (2008) Worldwide prevalence of anaemia 1993–2005: World Health Organization global database on anaemia. World Heal Organ. doi: 10.1017/S1368980008002401

2. Sysmex (2020) Advanced RBC parameters in the differential diagnosis and management of anaemia. Haematol white Pap. doi: www.sysmex-europe.com/whitepapers Sysmex

3. Jansen V (2019) Diagnosis of anemia-A synoptic overview and practical approach. Transfus Apher Sci 58:375-385

4. Means R, Brodsky R Diagnostic approach to anemia in adults. In: UpToDate. https://www.uptodate.com/contents/diagnostic-approach-to-anemia-in-adults. Accessed 20 Jan 2004

5. Schoorl M, Schoorl M, Linssen J, Villanueva MM, Velasco NoGuera JA, Martinez PH, Bartels PCM (2012) Efficacy of advanced discriminating algorithms for screening on iron-deficiency anemia and β-thalassemia trait: A multicenter evaluation. Am J Clin Pathol 138:300–304

6. Taher AT, Weatherall DJ, Cappellini MD (2018) Thalassaemia. Lancet 391:155–167

7. Urrechaga E, Borque L, Escanero JF (2011) The role of automated measurement of RBC subpopulations in differential diagnosis of microcytic anemia and β-thalassemia screening. Am J Clin Pathol 135:374–379

8. Benz E, Angelucci E Clinical manifestations and diagnosis of the thalassemias. In: UpToDate. https://www.uptodate.com/contents/clinical-manifestations-and-diagnosis-of-the-thalassemias.

9. Sirdah M, Tarazi I, Al Najjar E, Al Haddad R (2008) Evaluation of the diagnostic reliability of different RBC indices and formulas in the differentiation of the β-thalassaemia minor from iron deficiency in Palestinian population. Int J Lab Hematol 30:324–330

10. Ehsani MA, Shahgholi E, Rahiminejad MS, Seighali F, Rashidi A (2009) A new index for discrimination between iron deficiency anemia and beta-thalassemia minor: Results in 284 patients. Pakistan J Biol Sci 12:473–475

11. England J, Bain B, Fraser P (1973) Differentiation of iron deficiency from thalassaemia trait. Lancet 1514

12. Green R, King R (1989) A new red cell discriminant incorporating volume dispersion for differentiating iron deficiency anemia from thalassemia minor. Blood Cells 5:481-495

13. Mentzer WC (1973) Differentiation of iron deficiency from thalassaemia trait. Lancet 882

14. Ricerca B, Storti S, D'Onofrio G (1987) Differentiation of iron deficiency from thalassaemia trait: a new approach. Haematologica 72:409-413

15. Shine I, Lal S (1977) A strategy to detect beta-thalassaemia minor. Lancet (London, England) 1:692-694

16. Srivastava P, Bevington J (1973) Iron deficiency and/or thalassamia trait. Lancet 832

17. Urrechaga E, Hoffmann JJML (2017) Critical appraisal of discriminant formulas for distinguishing thalassemia from iron deficiency in patients with microcytic anemia. Clin Chem Lab Med 55:1582–1591

18. Jayabose S, Giamelli J, Levondoglu-Tugal O, Sandoval C, Ozkaynak F, Visintainer P (1999) Differentiating iron deficiency anemia from thalassemia minor by using an RDW-based index. J Ped Hematol Oncol 21:314

References

19. Janel A, Roszyk L, Rapatel C, Mareynat G, Berger MG, Serre-Sapin AF (2011) Proposal of a score combining red blood cell indices for early differentiation of beta-thalassemia minor from iron deficiency anemia. Hematology 16:123–127

20. Urrechaga E (2008) Discriminant value of % microcytic/% hypochromic ratio in the differential diagnosis of microcytic anemia. Clin Chem Lab Med 46:1752–1758

21. Hoffmann JJML, Urrechaga E, Aguirre U (2015) Discriminant indices for distinguishing thalassemia and iron deficiency in patients with microcytic anemia: A meta-analysis. Clin Chem Lab Med 53:1883–1894

22. Ullrich C, Wu A, Armsby C, Rieber S, Wingerter S, Brugnara C, Shapiro D, Bernstein H (2005) Screening Healthy Infants for Iron Deficiency Using Reticulocyte Hemoglobin Content. JAMA 294:924–930

23. Toki Y, Ikuta K, Kawahara Y, et al (2017) Reticulocyte hemoglobin equivalent as a potential marker for diagnosis of iron deficiency. Int J Hematol 106:116–125

24. Tiwari AK, Bhardwaj G, Arora D, Aggarwal G, Pabbi S, Dara RC, Sachdev R, Raizada A, Sethi M (2018) Applying newer parameter Ret-He (reticulocyte haemoglobin equivalent) to assess latent iron deficiency (LID) in blood donors-study at a tertiary care hospital in India. Vox Sang 113:639–646

25. Weimann A, Cremer M, Hernáiz-Driever P, Zimmermann M (2016) Delta-He, Ret-He and a new diagnostic plot for differential diagnosis and therapy monitoring of patients suffering from various disease-specific types of anemia. Clin Lab 62:667–677

26. Thomas L, Franck S, Messinger M, Linssen J, Thomé M, Thomas C (2005) Reticulocyte hemoglobin measurement - Comparison of two methods in the diagnosis of iron-restricted erythropoiesis. Clin Chem Lab Med 43:1193–1202

27. Thomas DW, Hinchliffe RF, Briggs C, Macdougall IC, Littlewood T, Cavill I (2013) Guideline for the laboratory diagnosis of functional iron deficiency. Br J Haematol 161:639–648

28. Thomas C, Kirschbaum A, Boehm D, Thomas L (2006) The Diagnostic Plot. Med Ocology 23:23-36

29. Thomas C, Thomas L (2002) Biochemical markers and hematologic indices in the diagnosis of functional iron deficiency. Clin Chem 48:1066–1076

30. Nivaggioni V, Bouriche L, Coito S, Le Floch AS, Ibrahim-Kosta M, Leonnet C, Arnoux I, Loosveld M (2020) Use of Sysmex XN-10 red blood cell parameters for screening of hereditary red blood cell diseases and iron deficiency anaemia. Int J Lab Hematol 42:697–704

31. Farias MG (2017) Advances in laboratory diagnosis of hereditary spherocytosis. Clin Chem Lab Med 55:944-948

32. Mentzer WC Hereditary spherocytosis. In: UpToDate. https://www.uptodate.com/contents/hereditary-spherocytosis.

33. King MJ, Garçon L, Hoyer JD, Iolascon A, Picard V, Stewart G, Bianchi P, Lee SH, Zanella A (2015) ICSH guidelines for the laboratory diagnosis of nonimmune hereditary red cell membrane disorders. Int J Lab Hematol 37:304–325

Mullier F, Lainey E, Fenneteau O, Da Costa L, Schillinger F, Bailly N, Cornet Y, Chatelain C, Dogne JM, Chatelain B (2011) Additional erythrocytic and reticulocytic parameters helpful for diagnosis of hereditary spherocytosis: Results of a multicentre study. Ann Hematol 90:759–768

35. Bobée V, Daliphard S, Schrapp A, Lahary A (2018) Screening of hereditary spherocytosis and pyruvate kinase deficiency by automated blood count using erythrocytic and reticulocytic parameters. Int J Lab Hematol 40:697–703

36. Sottiaux JY, Favresse J, Chevalier C, Chatelain B, Jacqmin H, Mullier F (2020) Evaluation of a hereditary spherocytosis screening algorithm by automated blood count using reticulocytes and erythrocytic parameters on the Sysmex XN-series. Int J Lab Hematol 42:e88–e91

37. Persijn L, Bonroy C, Mondelaers V, Vantilborgh A, Philippé J, Stove V (2012) Screening for hereditary spherocytosis in routine practice: Evaluation of a diagnostic algorithm with focus on non-splenectomised patients. Ann Hematol 91:301–302

References

38. Adam A-S, Cantinieaux B, Cotton F, Gulbis B (2020) Screening for hereditary spherocytosis: a new algorithm using Sysmex XN-9000 specific erythrocyte and reticulocyte parameters. Eur Hematol Assoc Libr Poster presentation.

39. Grace RF, Zanella A, Neufeld EJ, Morton DH, Eber S, Yaish H, Glader B (2015) Erythrocyte pyruvate kinase deficiency: 2015 status report. Am J Hematol 90:825–830

40. Beutler E, Gelbart T (2000) Estimating the prevalence of pyruvate kinase deficiency from the gene frequency in the general white population. Blood 95:3585–3588

41. Agarwal AM, Rets A (2020) Laboratory approach to investigation of anemia with a focus on pyruvate kinase deficiency. Int J Lab Hematol 42:107–112

42. Canu G, De Bonis M, Minucci A, Capoluongo E (2016) Red blood cell PK deficiency: An update of PK-LR gene mutation database. Blood Cells, Mol Dis 57:100–109

43. Kottke-Marchant K (2017) Diagnostic approach to microangiopathic hemolytic disorders. Int J Lab Hematol 39:69–75

44. Zini G, d'Onofrio G, Briggs C, Erber W, Jou JM, Lee SH, Mcfadden S, Vives-Corrons JL, Yutaka N, Lesesve JF (2012) ICSH recommendations for identification, diagnostic value, and quantitation of schistocytes. Int J Lab Hematol 34:107–116

45. Jiang M, Saigo K, Kumagai S, Imoto S, Kosaka Y, Matsumoto H, Fujimoto K (2001) Quantification of red blood cell fragmentation by automated haematology analyser XE-2100. Clin Lab Haematol 23:167–172

46. Lesesve JF, Speyer E, Perol JP (2015) Fragmented red cells reference range for the Sysmex XN®-series of automated blood cell counters. Int J Lab Hematol 37:583–587

47. Lesesve JF, Salignac S, Alla F, Defente M, Benbih M, Bordigoni P, Lecompte T (2004) Comparative Evaluation of Schistocyte Counting by an Automated Method and by Microscopic Determination. Am J Clin Pathol 121:739–745

48. Moiseev IS, Tsvetkova T, Aljurf M, et al (2019) Clinical and morphological practices in the diagnosis of transplant-associated microangiopathy: a study on behalf of Transplant Complications Working Party of the EBMT. Bone Marrow Transplant 54:1022–1028

49. Hantaweepant C, Sasijareonrat N, Chutvanichkul B, Karaketklang K, Chinthammitr Y (2020) Comparison between optical microscopy and the Sysmex XN-3000 for schistocyte determination in patients suspected of having schistocytosis. Heal Sci Reports 3:e138

50. Govindarajan S, Bhatia P, Dawman L, Tiewsoh K (2021) Usefulness of automated fragmented red blood cell percentage in the diagnosis of paediatric haemolytic uraemic syndrome. Int J Lab Hematol 43:40–43

51. Abe Y, Wada H, Yamada E, et al (2009) The effectiveness of measuring for fragmented red cells using an automated hematology analyzer in patients with thrombotic microangiopathy. Clin Appl Thromb 15:257–262

52. Chalvatzi K, Spiroglou S, Nikolaidou A, Diza E (2013) Evaluation of fragmented red cell (FRC) counting using Sysmex XE-5000 - Does hypochromia play a role? Int J Lab Hematol 35:193–199

53. Lesesve JF, Asnafi V, Braun F, Zini G (2012) Fragmented red blood cells automated measurement is a useful parameter to exclude schistocytes on the blood film. Int J Lab Hematol 34:566–576

54. Ory D, Kieffer D (2016) Red blood cell enzyme assays: (d)efficient? In: Crit. Apprais. Top. UZ Leuven. https://www.uzleuven.be/nl/laboratoriumgeneeskunde/critically-appraised-topics-cat 55. Urrechaga E, Borque L, Escanero JF (2009) Potential utility of the new sysmex XE 5000 red blood cell extended parameters in the study of disorders of iron metabolism. Clin Chem Lab Med 47:1411–1416

56. Pekelharing J, Hauss O, de Jonge R, Lokhoff J, Sodikromo J, Spaans M, Brouwer R, de Lathouder S, Hinzmann R (2010) Haematology reference intervals for established and novel parameters in healthy adults. Diagnostic Perspect 1:1–11