

Sputum smear microscopy in the diagnosis
of pulmonary tuberculosis:
What are the options anno 2014?

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Overview

- Introduction and History
- CAT questions
 - What are the current available and recommended staining methods for sputum smear microscopic examination in the diagnosis of tuberculosis?
 - What are the results of our own study, where three relevant staining methods are compared to each other?
- CAT answers
- General conclusions

Tuberculosis in a nutshell

- Described by Hippocrates (5th century BC) 'phtysis'
24 March 1982: Tubercle bacillus (Robert Koch)
- Infectious disease caused by *Mycobacterium tuberculosis*
- Air-born transmission
- Pulmonary VS extra-pulmonary
- Active VS latent
- Symptoms: chronic cough, fever, night sweats, weight loss
- Treatment: multiple antibiotics over a long period

Why Does TB Need Global Attention anno 2014?

- Deadliest infectious diseases affecting humans with yearly **2 million people who die** from tuberculosis
= 7% of all deaths
- Approximately **1/3** of the world population is infected with *M. tuberculosis*.
- **8-10 million new cases** of TB per year.
- **Leading cause of death** among people with HIV/AIDS.

➔ "global public health emergency"



Diagnosis of Pulmonary Tuberculosis

- Medical History
- Physical examination
- Sputum examination
 - Culture = reference
 - Microscopic examination
 - NAAT
- X-ray examination
- Tuberculine skin testing, gamma-interferon test

Diagnosis of Pulmonary Tuberculosis

Microscopic Sputum examination

- Minimum requirement of 5,000 to 10,000 CFU/mL
VS culture (10 tot 100 CFU/mL)
- Acid-fast staining procedure
 - Principle:
 - Wax mycol acid containing cell wall of Mycobacteria is impermeable to ordinary stainings
 - Heat softens the mycol wall and let allows the stain to enter
 - Phenol is soluble in lipids or waxes
 - Once stained it resists decolorisation by mineral acid (20% H₂SO₄)
 - Phenol-dye mixture is more soluble in the waxes of the cell wall than alcohol and acid
 - While the Mycobacteria retain the primary stain, the background is decolorized and takes up the counterstain
 - Ziehl-Neelsen VS fluorescent methods

Diagnosis of Pulmonary Tuberculosis

Microscopic Sputum examination

- Interpretation

Ziehl-Neelsen 1000x	Auramine 250x	Auramine 450x	Auramine 630x	Result
1-9/100gv	1-9/10gv	2-18/50gv	2-18/100gv	1+
1-9/10gv	1-9/10gv	4-36/10gv	2-18/10gv	2+
1-9/gv	10-90/gv	4-36/gv	2-18/gv	3+
>9/gv	>90/gv	>36/gv	>18/gv	4+

PART 1:

What are the most available staining techniques for the detection of acid-fast bacilli and what are the current recommendations?

Sputum Smear microscopy

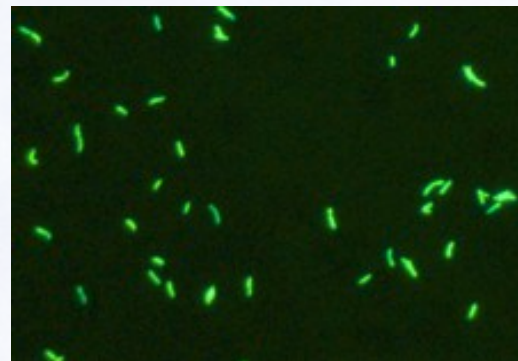
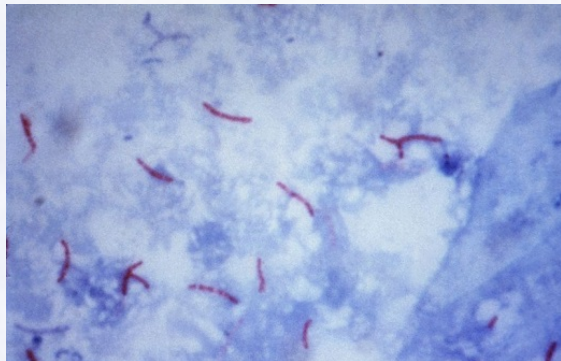
Conventional Light Microscopy

Koch – Ehrlich – Ziehl – Neelsen – Kinyoun

Fluorescence Microscopy

auramine O – auramine rhodamine – acridine orange

LED-Microscopy



Conventional Light Microscopy

- Robert Koch (1882)

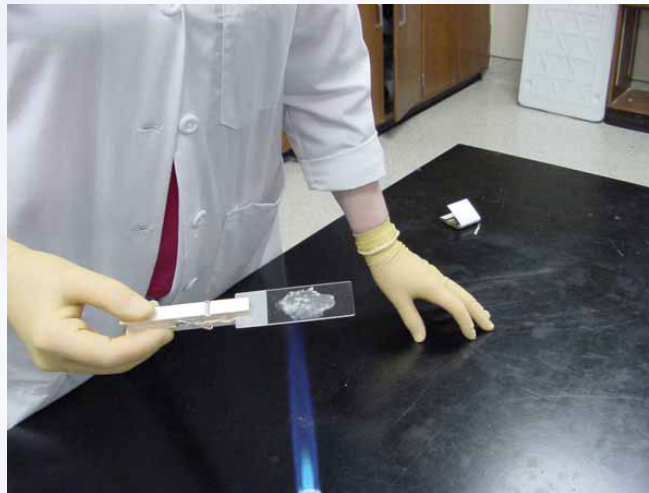
- Staining dried preparations in a weakly alkaline solution of methylene blue
- After 24h treatment with a solution of vesuvin (Bismarck's brown)
- Intense blue tubercles, with a brown background



Conventional Light Microscopy

- Paul Ehrlich (1882)

- Aniline VS methylene blue
- Shorter staining time: 15-30 minutes VS 24h
- Added 30% nitric acid and alcohol for decolorisation of surrounding tissues
- Counterstaining with yellow or blue dye
- Red tubercle bacilli more clearly than Koch's method
- Introduced 'heat-fixation': preparations 1h at 100-110°C or passing them three times through a Bunsen burner



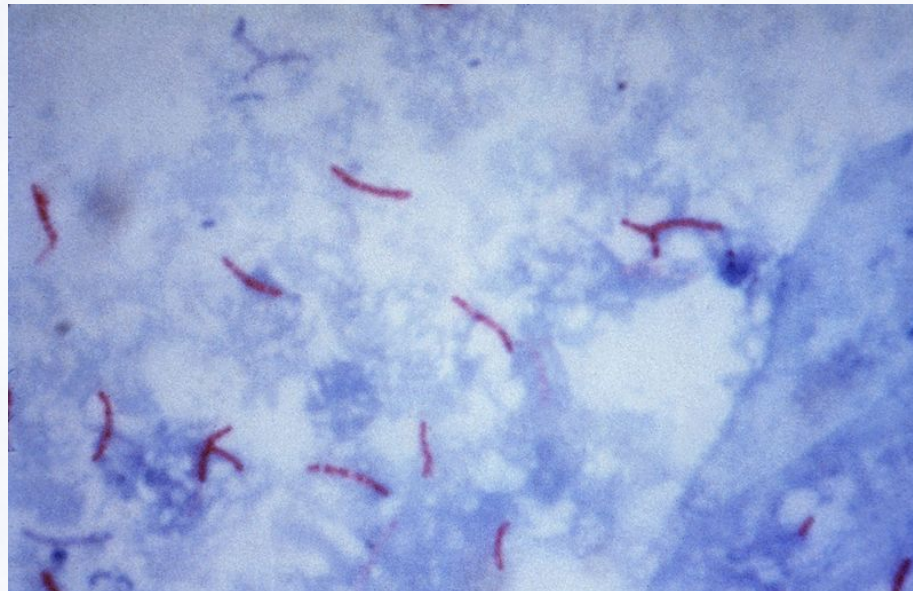
Conventional Light Microscopy

- Franz Ziehl (1857-1926)

Hot carbolic fuchsin VS aniline

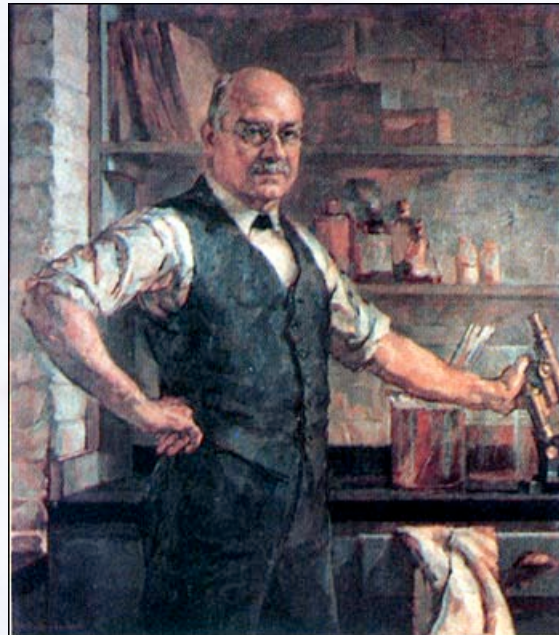
- Friedrich Neelsen (1854-1898)

Sulphuric acid VS nitric acid



Conventional Light Microscopy

- Joseph Kinyoun, 1914
 - Cold carbolic fuchsin VS heated carbolic fuchsin
 - 3% acid-alcohol for decolorizing
 - methylene blue or brilliant green for counterstaining



Conventional Light Microscopy

Ziehl-Neelsen

- Most applicable and available diagnostic tool of choice for diagnosis of TB in developing countries
- Rapid, inexpensive
- Excellent reported specificities: 96% - 100%
 - Highly specific in areas with high incidence
 - Luelmo F. What is the role of sputum microscopy in patients attending health facilities? Geneva: World Health Organisation, 2004:7-13
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- Variable reported sensitivities: 20% - 86%
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Conventional Light Microscopy

Ziehl-Neelsen: sensitivity

- *Variable reported sensitivities: 20% - 86%*
- Influence of other factors:
 - Prevalence/severity TB
 - Type of specimen
 - Method of processing (concentrated vs direct)
 - Method of centrifugation
 - Quality examination

Iademaro et al: ZN significant more sensitive than fluorochrome staining methods if prepared and interpreted following standard recommendations!

Conventional Light Microscopy

– Ziehl-Neelsen: sensitivity

- *Variable reported sensitivities: 20% - 86%*
- *Influence of other factors:*

lademarco et al: ZN significant more sensitive than fluorochrome staining methods if prepared and interpreted following standard recommendations!

In reality?

Sömovski et al:

- large proficiency testing for ZN microscopy
- 167 laboratories in the state NY, 91% used commercial staining kits
- Many unexpected errors:
 - Concentration carbol fuchsin
 - Time of staining and counterstaining
 - Concentration of acid alcohol for decolorization
 - Interpretation

Conventional Light Microscopy

Kinyoun

- Cold VS warm: no heating step required
- Anno 2014: ZN grossly replaced by Kinyoun
 - Less toxic
 - No need for sophisticated suction systems
- Diagnostic performance VS ‘heated’ ZN?
 - Lower reported sensitivities compared to classical ZN!
 - Sömovski, Collins, Allen, Slosarek, Gruft, Mathew

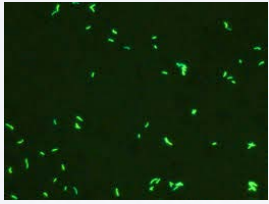


Conventional Light Microscopy

GUIDELINES

- Conventional light microscopy is not recommended in high-income countries for the diagnosis of TB
- If used so: classical Ziehl-Neelsen must be chosen over Kinyoun

NVMM, WHO, IDSA



Fluorescence Microscopy

- 1917, Kaiserling

Spontaneous fluorescence of *M. tuberculosis* under kristal-violet

- 1937, Hageman

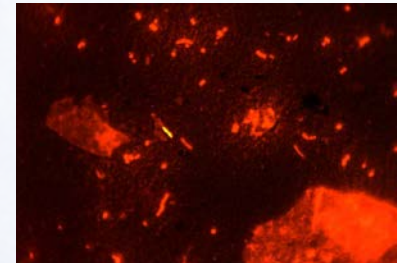
Auramine O or auramine-rhodamine as acid-fast fluorescent dye
Intense light source: halogen or high-pressure mercury vapour lamp

- 1982, Katila

Acridine orange VS auramine

- 1995, Smithwick

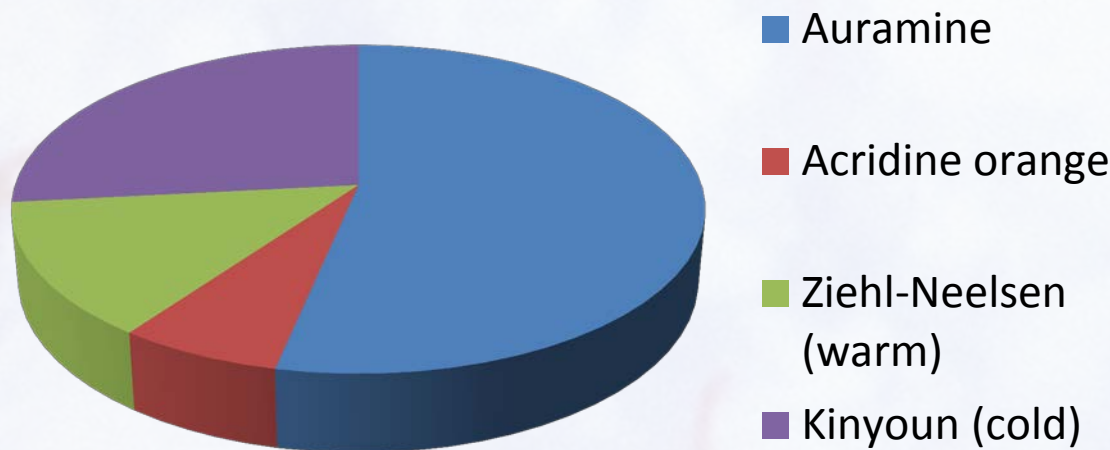
Introduction of phenol to accelerate dye penetration



Fluorescence Microscopy

- Most applied staining method for TB in high-income countries
- Belgium: (*questionary in 16 hospital-laboratories in Flanders*)

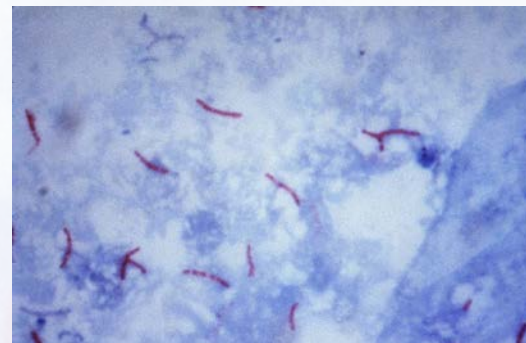
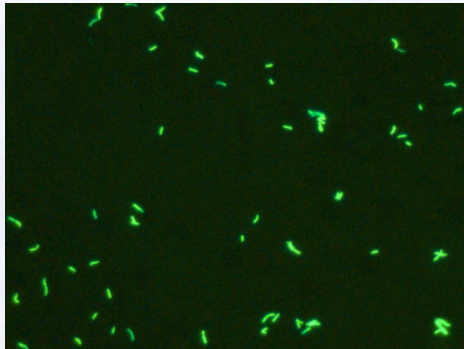
Staining Method for TB in 16 hospitals in Flanders



Fluorescence Microscopy

Practical advantages

- Use of a lower power objective lens (typically 25x)
VS conventional light microscopy (typically 100x)
 - Same area of slide more quickly and efficiently
 - 75% less time-consuming than CM
 - 15 minutes for CM VS approx 2,6 minutes for FM
- Easy and simple to recognise the acid-fast bacilli



Fluorescence Microscopy

Diagnostic performance

- Sensitivity

- Most studies result in better sensitivities of FM compared to CM
- Systematic review/thesis Henri, 2005
FM is more sensitive than CM
- Systematic review Steingart, 2009
52%-97%
FM 8-10% more sensitive than CM

- Specificity

- General concern related to less specific performance
 - ⇒ Guidelines recommend to confirm acid-fast bacilli by Ziehl-Neelsen (NVM)
 - ⇒ BUT:
 - Systematic review Steingart, 2009:
 - no decrease of specificity of FM compared to CM
 - den Hertog et al, 2013: retrospective study of 10,276 samples
 - no added value of confirming auramine-positive samples with Ziehl-Neelsen
 - Reanalysis of these samples have no impact on patient management and thus waste of resources

Fluorescence Microscopy

Practical disadvantages

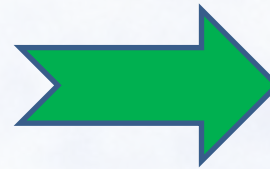
- High capital cost for conventional mercury vapour lamp microscopes
 - BUT: Kivighja(2003), Sohn(2009): proof of cost-effectivity of FM, even in low-income countries because of the high sensitivity and greater time efficiency
- Significant maintenance of the microscopes
- Limited life-span of the bulbs
- Need for a dark room, away from dusty environments
- Toxic exposure when broken

Fluorescence Microscopy

Global implementation?

PRO's:

- Diagnostic performance
- Easy to recognise the tubercles
- Less time-consuming
- Cost-efficient
- No need for confirmation with CM



HIGH-INCOME
COUNTRIES

CON's:

- High capital cost
- Significant maintenance
- Limited life-span bulbs
- Need for a dark room
- Away from dusty environments
- Toxic exposure when broken



~~LOW-INCOME
COUNTRIES~~

Next-generation Fluorescence Microscopy

Light-Emitting Diodes LED-microscopes

- *Martin, 2005*
 - *Described LED-microscopes used as excitatory light source for diagnostic fluorescence stains*
 - *“LED-microscopes could replace a mercury arc lamp for fluorescence microscopy”*



Next-generation Fluorescence Microscopy

Light-Emitting Diodes LED-microscopes

Practical advantages

- No need for a dark room
 - ➔ Improvement workflow
 - ➔ Maximum space utilisation in the lab
- Less maintenance required than FM
- Good durability and portability
- Less capital costs than FM
- Overall better cost-efficacy compared to CM:
 - Withlaw, 2011: **US\$2,10**CM VS **US\$1,63** LED
 - Xia, 2014: **US\$2,20** (+/-0,58)CM VS **US\$1,97** (+/-0,71)LED $p < 0,05$

Next-generation Fluorescence Microscopy

Light-Emitting Diodes LED-microscopes

Diagnostic Performance

- Meta-analysis WHO expert group, 2009:
 - Sensitivity of LED is significantly better compared to Ziehl-Neelsen: 6% (95%CI, 0,1-13%) with similar specificities
 - Sensitivity and specificity of LED is significantly better compared to FM: 5% (95% CI,0-11%) resp. 1% (95% CI,-0,7-3%)
- Most more recent studies confirm these findings

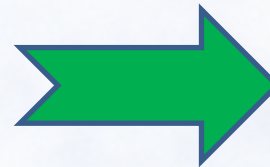
	LED-FM	FM	CM	P-value
Cuevas (2011)	72,8%		65,8%	<0,001
Bonnet (2011)	73,2%		72,0%	=0,32
Khatun (2011)	95,4%	68,1%	56,1%	<0,001
Bhala (2013)	83,1%	82,2%	78,0%	
Marzouk (2013)	82,2%	82,2%		
Albert (2010)	+5,6-9,4%			

Next-generation Fluorescence Microscopy

Global implementation?

PRO's:

- Diagnostic performance
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HIGH-INCOME
COUNTRIES

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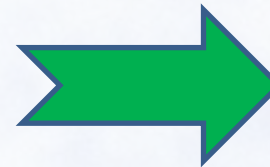
~~LOW-INCOME
COUNTRIES~~

Next-generation Fluorescence Microscopy

Global implementation?

PRO's:

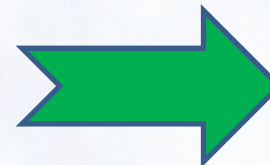
- Diagnostic performance
- Easy to recognise the tubercles
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HIGH-INCOME
COUNTRIES

CON's:

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LOW-INCOME
COUNTRIES

Next-generation Fluorescence Microscopy

Guidelines

- FM be replaced by LED-microscopy in all settings where FM is currently used
- LED microscopy be phased in as an alternative to CM in both high- and low-income laboratories

WHO2011

PART 2:

Field study:

A small prospective study in order to compare diagnostic performance of three relevant staining techniques for the detection of acid-fast bacilli: Ziehl-Neelsen, auramine O and acridine orange.

Study

- Many studies evaluated performance of CM and FM, compared to each other
- FM: most of them related to auramine
- AZ Sint-Jan Brugge:
 - Acridine Orange staining:
 - ➔ scarce literature concerning diagnostic performance
 - Katila (1982) and Smithwick (1995): diagnostic performance Acridine Orange comparable to Auramine staining
 - Kalich (1989): Acridine orange outperforms Auramine O
 - Narayan (2012): superior sensitivity of Acridine Orange compared to Auramine O

Material and Methods

- Prospective study AZ Sint-Jan, Bruges, AZ Zeno, Knokke-Heist/Blankenberge and UMC Saint Pierre/Bordet Instituut Brussels
- 200 routine respiratory samples, patients with clinical suspected TB
- After decontamination, three smear slides were prepared.
- Smears were stained by standard recommendations with Ziehl-Neelsen, auramine O and acridine orange
- Stained slides were examined following standard recommendation by experienced laboratory staff/clinical biologists.

Results

- 10,3% positive cultures = reference method

	Ziehl-Neelsen	Auramine O	Acridine Orange
Sensitivity	66,67% (95CI 43,04-85,35)	66,67% (95CI 43,04-85,35)	61,90% (95CI 38,45-81,84)
Specificity	97,55% (95CI 93,83-99,31)	100% (95CI 97,73-100)	99,38% (95CI 96,6-99,9)
PPV	77,78% (95CI 52,36-93,45)	100% (95CI 76,66-100)	92,86% (95CI 66,06-98,81)
NPV	95,78% (95CI 91,5-98,28)	95,86 (95CI 91,65-98,31)	95,27% (95CI 90,88-97,93)

General Answers and Conclusions

- Sputum smear microscopy remains the most important diagnostic tool for detecting acid-fast bacilli
- Guidelines and other published data:
 - LED fluorescence microscopy gains importance
 - Ease in use/interpretation
 - Cost-effectivity
 - Diagnostic performance
 - ➔ Reference staining method in high-income countries
 - ➔ Not globally implemented yet
 - Conventional light microscopy: Ziehl-Neelsen/Kinyoun loses importance
 - More difficult to interpret
 - More expensive
 - Less performant
 - However: not confirmed by our field study => if prepared and interpreted following standard recommendations equal sensitivities? (Iademarco and own results)
 - ➔ Still the most known, available and applied staining method in developing countries
 - ➔ More and more abandoned in high income countries

Did's and To Do's

- Field study: overall good performance of the three staining methods. Too small for adequate conclusions concerning comparison between the methods.
 - Only a limited of studies have been performed evaluating the diagnostic performance of Acridine Orange staining for the detection of acid-fast bacilli.
-
- ➔ Because of good performance of the field study and it's succesfull validation, the Acridine Orange staining is implemented in our lab for the detection of acid-fast bacilli.
 - ➔ Follow-up literature concerning diagnostic performance of Acridine Orange in the detection of acid-fast bacilli.

Literature

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